Cystic fibrosis

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Abstract | Cystic fibrosis is an autosomal recessive, monogenetic disorder caused by mutations in the cystic fibrosis transmembrane conductance regulator (*CFTR*) gene. The gene defect was first described 25 years ago and much progress has been made since then in our understanding of how *CFTR* mutations cause disease and how this can be addressed therapeutically. **CFTR** is a transmembrane protein that transports ions across the surface of epithelial cells. CFTR dysfunction affects many organs; however, lung disease is responsible for the vast majority of morbidity and mortality in patients with cystic fibrosis. Prenatal diagnostics, newborn screening and new treatment algorithms are changing the incidence and the prevalence of the disease. Until recently, the standard of care in cystic fibrosis treatment focused on preventing and treating complications of the disease; now, novel treatment strategies directly targeting the ion channel abnormality are becoming available and it will be important to evaluate how these treatments affect disease progression and the quality of life of patients. In this Primer, we summarize the current knowledge, and provide an outlook on how cystic fibrosis clinical care and research will be affected by new knowledge and therapeutic options in the near future. For an illustrated summary of this Primer, visit: <u>http://go.nature.com/4VrefN</u>

Cystic fibrosis is an autosomal recessive disease caused by mutations in the cystic fibrosis transmembrane conductance regulator (CFTR) gene. Close to 2,000 mutations in this gene have so far been described (see the Cystic Fibrosis Mutation Database), although fewer than 150 are known to be disease causing (see the Clinical and Functional Translation of CFTR (CFTR2) website). Although cystic fibrosis is a monogenetic disease, its phenotypic variability is substantial - as shown by the broad range of disease severity observed in patients with the same genotype¹. The cystic fibrosis phenotype (BOX 1) is characterized by progressive lung disease, exocrine pancreatic insufficiency that results in gastrointestinal malabsorption, intestinal abnormalities that result in malnutrition, impaired growth, and a variety of other manifestations, including sinusitis and diabetes. CFTR primarily functions as a chloride channel that transports ions across the apical membrane of epithelial cells throughout the body, but has other functions, including bicarbonate secretion and inhibition of sodium transport, which are important for the pathophysiology of CFTR deficiency and dysfunction. Mutations in CFTR are grouped in classes that reflect their functional consequences; those leading to loss of CFTR expression on the cell surface or loss of its function are generally 'severe' mutations associated with a phenotype of both lung disease and pancreatic insufficiency. Mutations with residual CFTR function are often associated with preserved pancreatic function;

some individuals show single organ manifestations, such a congenital bilateral absence of the vas deferens in the male reproductive tract², *CFTR* mutations have also been described in patients with cystic fibrosis-like organ manifestations — including pancreatitis, sinusitis or 'idiopathic' bronchiectasis (widening of the airways) — and the threshold of CFTR function needed to prevent disease varies between different organs^{3,4}.

The discovery of the cystic fibrosis gene defect in 1989 has resulted in a better understanding of disease pathophysiology, but only in the past few years has this information led to targeted therapies that address the underlying cellular defect⁵⁻⁷. Important advances in addressing all aspects of the disease have been made over the past two decades and the prognosis of patients with cystic fibrosis is constantly improving. The development of clinical trial networks has enabled rapid testing of putative treatments through training of research staff, establishment of standard operating procedures and reference laboratories. Strong partnerships have also been developed between government agencies, academic centres, voluntary health organizations, individuals and families with cystic fibrosis. Although these advances can mainly be attributed to improvements in symptom-directed therapy of downstream complications, it is expected that upstream CFTR-directed therapy will further improve patients' longevity and quality of life. In this Primer, we summarize the current understanding about how CFTR mutations cause disease and

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> we outline current diagnostic strategies and therapeutic interventions addressing disease manifestations. We also describe the epidemiology of cystic fibrosis and how cystic fibrosis and its treatment affect patients' quality of life. Lastly, we discuss how cystic fibrosis care and treatment is expected to change in future years.

Epidemiology

Changing diagnostic criteria and methods as well as improvements in clinical outcome have influenced the epidemiology of cystic fibrosis. Estimates of disease incidence are around 1 in 3,000 live births in persons of northern European descent^{8,9}, with Ireland having the highest incidence at 1 in 1,400 live births¹⁰. Incidence varies according to race and ethnicity; only 1 in 4,000 to 10,000 Latin Americans and 1 in 15,000 to 20,000 African Americans have cystic fibrosis, with even lower incidence rates in people of Asian background⁹. These estimates are based on information from western countries — epidemiological data are missing for large regions of the world, including the Middle East, Asia and Africa. Importantly, some small populations in eastern Europe have very high incidence rates, specifically Albania, where the incidence

Box 1 | Phenotypes of cystic fibrosis

Classic cystic fibrosis

- Chronic sinusitis
- Pancreatic insufficiency
- Diabetes
- Obstructive lung disease from infancy with chronic bacterial infection
- Severe liver disease (5–10% of patients)
- Meconium ileus (15–20% of patients)
- Congenital bilateral absence of vas deference (males)
- High sweat chloride concentrations

Nonclassic cystic fibrosis

- Chronic sinusitis
- Pancreatic sufficiency
- Pancreatitis (5–20%)
- Obstructive lung disease (variable onset)
- Congenital bilateral absence of vas deference (males)
- Lower sweat chloride concentrations than classic
- cystic fibrosis

was noted to be 1 in 555 (REF. 11). This high incidence is also reflected in data noting very high incidence in Albanian immigrants to northern Italy¹². The introduction of prenatal genetic screening in western countries seems to correlate with decreasing incidence in some countries¹³. Although the incidence is decreasing, data from registries suggest that the prevalence is increasing because of improvements in survival¹⁴⁻¹⁶ (FIG. 1).

Recent analyses in the United States have shown that cystic fibrosis survival improved in the period from 2000 to 2010 at a rate of 1.8% per year (95% CI, 0.5-2.7%) and that the projected median survival of children born today is 56 years (95% CI, 54-58 years) if the mortality rate continues to decrease at this rate¹⁵. However, median age of death is still in the mid-twenties to early thirties¹⁶. These data mirror earlier findings from the United Kingdom¹⁷. Consistent with improved survival, the diagnosis of cystic fibrosis has moved to early in life with universal newborn screening in many countries. In 2010, more than half (58%) of the people with cystic fibrosis in the United States were diagnosed by newborn screening compared with only 8% of those diagnosed in 2000. Several countries are now reporting that >50% of their cystic fibrosis population is >18 years of age18,19. Not only has the patient population aged but also an increasing number of people with mild phenotypes of cystic fibrosis have been diagnosed on the basis of advances in genotyping of CFTR mutations, which contributes to, but does not fully account for, the increase in survival. In patients 40 years and older who were diagnosed after the age of 15 years, the median age of diagnosis has been reported to be 48.8 years (range, 24 to 72.8 years)²⁰; these individuals are much more likely to have a nonclassic phenotype (see below)^{21,22}. Survival is improving, even in patients with severe lung disease (defined as a forced expired volume in 1 second (FEV,) <30% of predicted)23, with median survival in those patients not receiving lung transplants increasing from 1.2 to 5.3 years in the period from 1991 to 2002.

Key questions in cystic fibrosis epidemiology remain unanswered. The first question relates to the exact effect of the almost universal use of newborn screening; newborn screening could enable children to maintain their lung health into adulthood. The next key question is how the advent of CFTR-directed therapies such as ivacaftor^{24,25} will change the long-term morbidity and mortality. In the future, incidence might continue to fall, partly because of prenatal counselling and partly because of mixing of populations with differing incidence of *CFTR* mutations. However, prevalence could continue to increase as a consequence of the initiation of new therapies prior to end-organ injury.

Mechanisms/pathophysiology CFTR protein and genetic mutations

Cystic fibrosis is caused by gene mutations in *CFTR* on the long arm of chromosome 7 (REFS 5,26). This gene is a unique member of the ATP-binding cassette (ABC) or traffic ATPase family of genes^{27,28}, which carry a regulatory domain that is actively phosphorylated^{29,30}. CFTR primarily functions as an apical anion channel of



Figure 1 | **Cystic fibrosis in the United States. a** | The number of patients reported in the annual reports of the Cystic Fibrosis Foundation in 5-year intervals from 1998 to 2013 are shown. The figure notes a fairly substantial increase in the number of people living with cystic fibrosis in the US Cystic Fibrosis Foundation Patient Registry, which is probably attributable to improved survival of patients during this 15-year period and not merely newly diagnosed patients. **b** | The numbers of newly diagnosed patients in each of the given years over time are shown.

chloride and bicarbonate, rather than as an active pump. Similarly to other members of the ABC protein family, it includes two nucleotide-binding domains (NBDs) encoding sites capable of binding and hydrolysing ATP (Walker A and B motifs) and membrane-spanning domains that function as the ion channel pore through the plasma membrane (FIG. 2).



Figure 2 | **Structure of CFTR. a** | Linear structure of cystic fibrosis transmembrane conductance regulator (CFTR) is shown. **b** | The CFTR protein is comprised of two, six span membrane-bound regions each connected to a nuclear binding domain (NBD; NBD1 and NBD2). NBDs bind ATP, as well as a regulatory (R) domain that consists of many charged amino acids. The channel opens when its R domain is phosphorylated by protein kinase A and when ATP is bound at the NBDs. MSD, membrane spanning domain.

CFTR mutations can reduce channel number, function or both, and can vary in severity and occur through a variety of cellular mechanisms. The relative severity and completeness of each genetic defect has a major influence on the manifestations and the severity of disease (that is, the cystic fibrosis phenotype), although genetic modifiers and environmental factors also have a role (see the Clinical and Functional Translation of CFTR (CFTR2) website)^{8,31}. The presence of at least one CFTR allele that is partially active can vastly improve clinical outcome; this is also evident from lower concentrations of sweat chloride or higher likelihood of pancreatic sufficiency compared with those that have no functional CFTR. When two mild or 'variable' mutations are present, or one mutation has sufficient function, atypical forms of cystic fibrosis can occur, such as congenital absence of the vas deferens³², 'idiopathic' pancreatitis^{33,34} or very late-onset respiratory disease without other characteristic features of cystic fibrosis³²⁻³⁴. A global catalogue of the phenotypes found in reported patients (patients who have been included in registries and report to the database) is available online (see the Clinical and Functional Translation of CFTR (CFTR2) website)³⁵ and can provide diagnostic and prognostic guidance to the management of individual patients.

The classification of CFTR alleles into molecular classes helps to simplify our understanding of the cellular defect (TABLE 1); however, it is now clear that many mutations have more than one feature³⁶. This has led to a multinational effort to standardize the characterization of the mutations, particularly with regard to their response to therapeutics (that is, the 'theratype'), and to publicize the information through open data repositories³⁵. The function of the CFTR protein can be reduced by disordered regulation and gating, which causes diminished ATP binding and hydrolysis (class III alleles) or defective chloride conductance (class IV alleles). Reduced channel number can be conferred by major disruptions of the CFTR gene (insertions, deletions and premature termination codons; each representative of a class I allele), aberrant splicing that reduces or eliminates full-length and stable mRNA transcripts (non-canonical and canonical splicing mutations, respectively; class V alleles), mutations that reduce surface stability (class VI alleles) or misfolding mutations that destabilize the CFTR protein and that subject it to premature degradation by endoplasmic reticulum-associated degradation, disrupting its normal localization to the plasma membrane (class II alleles). Class II mutations include the most common CFTR allele, deletion of the phenylalanine at position 508 (also known as F508del or c.1521_1523delCTT).

The severity of disease depends on whether a molecular defect is complete, in addition to other factors unrelated to the CFTR protein itself, such as modifier genes or environment. Mutations that affect chloride conductance are frequently only mild in severity; similarly, some class II and class VI alleles only partially disrupt protein stability, and non-canonical splice mutations are — by definition — partial, owing to the presence of normally spliced transcripts. Some alleles also show more than one

Molecular classification	Functional abnormality	Molecular consequence	Primary therapy type	Example mutations	
Class I	Decreased number	No functional protein produced	Translational readthrough*	G542X, 394delTT and 3905insT	
Class II	Decreased number	Absent or diminished protein processing	Corrector*	F508del, N1303K and A455E	
Class III	Decreased	Defective gating	Potontiator*		
Class III	function [§]	Delective gating	FOLEIILIALOI	and F508del [‡]	
Class IV	Decreased function	Decreased conductance	Potentiator*	R347H, R334W and R117H [‡]	
Class V	Decreased number	Abnormal splicing	Splice repair [∥]	621+1G>T,	
		(canonical (complete) or non-canonical (partial))	Potentiator	3849 + 10kbC>1 and R117H+5T	
Class VI	Decreased	Decreased cell surface	Cell surface stabilizer	S1455X, L1399X and F508del [‡]	
	number	stability	Potentiator		

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*Therapy type that has shown activity in one or more clinical trials. [‡]Secondary abnormality. [§]Open channel probability. ^{IP}Otential therapy type.

abnormality, adding further complexity. Although the main defect that results from F508del is destabilized protein folding due to altered NBD1 stability, which subjects the protein to degradation in the proteasome, the protein also alters CFTR gating and cell surface residence time³⁷. Indeed, F508del is at a key position within NBD1 such that it causes destabilization; furthermore, the deletion affects interdomain assembly and the communication of conformational changes to the transmembrane domains that are necessary for channel activation^{38,39}.

In addition to CFTR mutations, disease manifestations and progression are influenced by non-CFTR gene modifiers and environmental factors^{31,40-43}. For example, multiple genes that encode apical plasma membrane proteins found near CFTR lead to an increased risk of meconium ileus (a condition whereby the bowel is obstructed by viscous secretions) in the newborn^{42,44,45}. The meconium ileus risk alleles in solute carrier family 26 member 9 (SLC26A9; which mediates processes near CFTR), SLC9A3 and SLC6A14 are pleiotropic and influence other cystic fibrosis comorbidities in early life, including earlier development of lung damage and acquisition of Pseudomonas aeruginosa infection at a younger age46. A large and ongoing genome-wide association study and linkage study of 3,600 patients with cystic fibrosis reported a strong association between lung disease severity and loci on chromosome 11 and chromosome 20 (REF. 47).

In several analyses from an international twin and sibling study, genetic modifiers independent of CFTR are estimated to account for up to 50% of the variation in lung function in patients with cystic fibrosis, with the remainder attributed to environmental exposures and stochastic effects^{48,49}. Genetic variations in innate immune molecules (for example, mannose-binding lectin 2 and chloride channel accessory 4)⁵⁰ and cytokines (for example, transforming growth factor β 1 (TGF β 1) and tumour necrosis factor (TNF)) have been examined in many studies for their influence on clinical phenotype^{41,51,52}. The results have not been consistent in all

populations studied, which probably reflects differences in the populations studied (in terms of population size and the distribution of *CFTR* mutation status). These studies have identified potential modifiers that can be further assessed in the large cohorts that have been established for genome-wide association studies. For example, the risk for the development of cystic fibrosis-related diabetes has recently been shown to be influenced by modifier genes that include variants at *SLC26A9* and at four susceptibility loci for type 2 diabetes mellitus⁴³.

Environmental factors that have been shown to influence disease progression include exposure to environmental factors or toxins (for example, second-hand smoke, pollutants, climate and exposure to microorganisms), access to specialized care, adherence to treatment and variation in clinical practice⁵³⁻⁵⁶. Importantly, tissue-specific differences are evident in the susceptibility to CFTR mutations, non-CFTR gene modifiers and environmental factors. For example, specific CFTR gene mutations strongly affect pancreatic function (phenotype)⁵⁷, but environmental and other non-genetic factors have a greater influence on the pulmonary phenotype. The combination of large cohorts with the reduced costs and the enhanced processing of vast genetic data in the coming years are likely to provide understanding of the complex relationships between CFTR, non-CFTR gene modifiers and environmental factors on disease manifestations in cystic fibrosis.

Airway pathophysiology

Cystic fibrosis affects the function of epithelial tissues in which CFTR is highly expressed; in particular, glandular epithelia. The disease primarily manifests in the lungs, pancreas, gastrointestinal tract, vas deferens and sweat glands, although airway disease is the main cause of morbidity and mortality. In the lungs, cystic fibrosis results in mucus accumulation that compromises the airway lumen and that contributes to obstructive pulmonary disease (FIG. 3). Submucosal gland hyperplasia and thickened mucus secretions are also prominent.



Figure 3 | **The mucociliary transport defect in cystic fibrosis. a** | In the healthy state, adequate airway surface homeostasis enables effective transport of mucus extruding from the airway surface goblet cells and submucosal glands. Appropriate bicarbonate and pH regulation enable normal mucus to form, which facilitates the formation of a two-layer gel that optimizes mucociliary clearance and airway defence. **b** | Depletion of the airway surface liquid occurs through the absence of cystic fibrosis transmembrane conductance regulator (CFTR)-mediated fluid secretion accompanied by tonic fluid absorption via the epithelial sodium channel (ENaC; inset). CFTR-dependent liquid desiccation decreases airway surface liquid (ASL) depth, including the periciliary layer, ultimately contributing to mucus stasis. Decreased bicarbonate transport contributes to an acidic pH. Abnormally adherent mucus also emerges from the glands in cystic fibrosis and can become fixed to the gland orifice or to the originating goblet cells. These events contribute to a proinflammatory airway environment that further accelerates pathogenesis.

Airway disease is thought to begin in the small airways⁷. Development of bronchiectasis leads to irreversible changes that encourage continued infection and that accelerate disease pathogenesis.

Despite decades of research, the understanding of the origins of airway pathogenesis remains incomplete and continues to evolve. Several key manifestations include delayed mucociliary clearance through airway surface liquid depletion, abnormalities of the physical properties and adhesion of mucus, and a predisposition to infection owing to abnormal mucosal defences^{58,59}. Dysregulated inflammation intrinsic to the *CFTR* defect is also apparent. These processes initiate and perpetuate a cycle of destruction that ultimately results in irreversible lung injury, bronchiectasis and respiratory failure^{58,59}.

Loss of apical CFTR leads to reduced chloride and bicarbonate secretion⁶⁰. Given that the release of water and electrolytes onto the airway surface is mostly driven by CFTR-dependent fluid secretion through both the glands and the surface epithelia, CFTR deficiency leads to diminished airway surface hydration, which can impair mucociliary transport in itself⁶⁰. CFTR has also been shown to regulate the activity of the epithelial sodium channel (ENaC; also known as the amiloridesensitive sodium channel), which is also activated by cleavage events conferred by free proteases such as prostasin and neutrophil elastase that are enriched in the inflamed airway⁶¹⁻⁶³. CFTR decrement also confers unopposed ENaC-dependent sodium and water absorption, which exacerbates airway surface liquid depletion⁶⁴. Moreover, the periciliary layer, which is a mucin gel layer between the cell surface and the mucus layer (FIG. 3), is sensitive to the osmolar forces of the overlying mucus; as the overlying mucins become concentrated in the absence of adequate fluid transport, the periciliary layer collapses and failure of mucociliary transport ensues⁶⁵. However, this view is not without controversy. Specifically, ENaC hyperactivity has been questioned owing to the absence of sodium hyperabsorption from airway mucosa in newborn pigs with cystic fibrosis (together with evidence of normal airway surface liquid depth)⁶⁶, the lack of heightened amiloride-sensitive currents in rat models67 and the finding that elevated amiloride-sensitive currents can be increased by reduced CFTR-dependent transepithelial resistance in primary human airway epithelial monolayers derived from patients with cystic fibrosis68. Even in the presence of adequate airway hydration, mucus abnormalities can adversely affect mucociliary transport⁶⁹, and mucus adhesion can occur at the gland outlet even under submerged conditions, which suggests multifactorial causes to delayed mucociliary clearance70.

Mucus and glandular epithelium abnormalities

Hyperviscous respiratory secretions obstruct small and medium airways, leading to marked failure of mucociliary clearance that can be macroscopically verified by radioligand imaging⁷¹. A primary biochemical defect in mucus composition has been explored but is not well established as a fundamental cause of the disease, although enzymatically driven events that lead to the release of intestinal mucins have been identified⁷². Mucus includes a complex range of extracellular proteins, which are found in high concentrations in the airway lumen⁷³.

In extrapulmonary organs (for example, the pancreas and the liver), marked ductular obstruction is observed in the absence of polymicrobial infection, which enables direct studies of the relationship between CFTR and

mucus formation to be carried out. An emerging idea implicates defective bicarbonate transport as a mediator of hyperviscosity and mucosal adhesion in cystic fibrosis⁷⁴. In this model, exocrine mucus — which is highly negative in charge - is produced by acinar and other epithelial cells. This mucus binds calcium ions, which condenses the mucus and shields the negative repulsive force between sulphates and other anionic groups on constituent mucins74. Bicarbonate secretion via CFTR chelates calcium and permits mucinous expansion and a viscoelastic state that is compatible with physiological clearance. Failure of bicarbonate release is hypothesized to result in defective mucin expansion, leading to hyperviscous secretion with abnormally adherent properties; that is, the mucus is tightly bound to the epithelial surface and is difficult to mobilize. Evidence of excessive mucus viscosity and adhesion that depends on bicarbonate secretion has been observed in the intestines of mouse models of cystic fibrosis^{75,76} and excised airways of model pigs⁶⁹. Notably, CFTR is highly expressed in the glandular epithelium, where it activates fluid and electrolyte secretion68,70; CFTR-dependent anion transport is also crucial for the release of elastic mucus from the gland duct, even under submerged (hydrated) conditions, which indicates its importance to normal mucus maturation and transport70. As only large animal models of cystic fibrosis (for example, pig, ferret and, to a lesser extent, rat) have prominent glandular expression, this might explain why mouse models are minimally affected by lung disease67,77,78.

Defects in airway defence

Although CFTR mainly functions as an anion transporter, it also regulates numerous processes, including fundamental aspects of airway defence and inflammatory cell function. CFTR is situated within membrane complexes in close proximity to several integral membrane proteins, including other ion channels. In addition to ENaC, CFTR can directly or indirectly regulate anion secretion through other chloride channels, such as transmembrane member 16A (TMEM16A; also known as anoctamin 1), or can contribute to airway pH regulation through chloride exchangers, including anion exchange protein 2 (REFS 79,80). The absence of bicarbonate secretion leads to an acidic pH airway surface liquid in cystic fibrosis, which has been reported as a possible cause of defective bacterial killing by the highly pH-sensitive innate defensins⁸¹. CFTR also has a direct effect on neutrophil killing, as it affects degranulation by interfering with granule trafficking⁸². Dysfunctional macrophages might also be biased towards a proinflammatory response⁸³. Proteomic and transcriptomic analyses show hundreds of cellular gene products that directly bind to or that are regulated by CFTR. Accordingly, additional effects of CFTR are likely to emerge, partly fostered by studies using agents that specifically activate the protein⁸⁴.

Inflammation in lung disease. Whether infection is required to cause airway inflammation in cystic fibrosis remains under debate. Post-mortem studies have reported that babies with cystic fibrosis who died from

meconium ileus had normal (or near to normal) airway epithelium and no signs of inflammation or infection⁸⁵. However, animal models have suggested that abnormalities of mucus can be proinflammatory in the absence of infection⁸⁶, and endoplasmatic reticulum-associated protein degradation induced by misprocessed F508del CFTR might also provide a stimulus for a heightened inflammatory response⁸⁷. Despite this controversy, what is clear is that infection exaggerates the inflammatory milieu. Excessive neutrophil-dominated inflammation is observed in the absence of infection⁸⁸, and neutrophil elastase activity (a marker of airway inflammation) in bronchoalveolar lavage fluid in infants (3 months of age) was associated with early bronchiectasis at 12 months and 3 years of age in children with cystic fibrosis⁸⁹.

Factors released in chronic neutrophilic inflammation in cystic fibrosis can markedly reduce airway surface liquid height⁹⁰. As well as aggravating airway dehydration, the imbalance between neutrophil elastase, other proteolytic enzymes (derived from inflammatory cells or bacteria) and anti-proteases results in exaggerated tissue damage^{91,92}. Oxidative stress and persisting airway inflammation might be associated with local airway deficiency in glutathione (which normally protects from reactive oxygen species)^{93,94}. Over time, a vicious cycle of reduced mucus clearance, neutrophil-dominated inflammation and bacterial infection damages the airways⁹⁵.

Lung infection. Airway pathogens that are most commonly detected include *P. aeruginosa*, *Staphylococcus aureus*, and Aspergillus species. *P. aeruginosa* infection has been associated with increased mortality, frequent exacerbations, rapid decline in lung function and heightened inflammation in patients with cystic fibrosis, which has fuelled programmes of eradication over the past two decades^{96–98}. Viral infection is also a common cause of exacerbations in people with cystic fibrosis^{99,100} — although these patients are not more susceptible to viral infection, the effect is greater¹⁰¹. Fungi, including Aspergillus species, are also increasingly recognized as pathogens in cystic fibrosis and are associated with an increased rate of pulmonary exacerbations¹⁰².

With increasing age, infections with other bacteria, including Achromobacter xylosoxidans, Stenotrophomas maltophilia and Burkholderia cepacia complex, become increasingly common¹⁶. The prevalence of each of these pathogens varies between cystic fibrosis populations, although they generally occur in <15% of patients and rates increase with disease severity. Methicillin-resistant S. aureus (MRSA) is an increasing threat that occurs in up to 30% of patients presenting at some US centres^{16,103}, although the rates are lower in Europe. In the late 1980s, epidemic strains of Burkholderia cenocepacia were associated with clear evidence of cross-infection, rapid clinical deterioration in many, and poor outcomes following lung transplantation¹⁰⁴. Both MRSA and B. cenocepacia infection in patients with cystic fibrosis have been reported to have adverse effects on prognosis^{103,105}. Although P. aeruginosa was previously thought to be acquired from the environment, sharing of strains (patients with

Box 2 | Conventional diagnostic criteria for cystic fibrosis

Individuals must have at least one clinical feature:

- Meconium ileus
- Diarrhoea and failure to thrive
- Recurrent respiratory infections
- Nasal polyps
- Rectal prolapse
- Male infertility
- Electrolyte depletion

or a diagnosis of cystic fibrosis in a sibling and/or a positive newborn screening test, plus laboratory evidence of an abnormality in the cystic fibrosis transmembrane conductance regulator (*CFTR*) gene or protein:

- Chloride channel dysfunction (positive sweat test or abnormal transepithelial potential difference)
- Known disease-causing mutations on chromosome 7 in trans.

the same strains) has been shown to be common over the past 15 years, with some strains being associated with adverse clinical outcomes¹⁰⁶⁻¹⁰⁸. In some settings, strong evidence supports person-to-person spread of shared strains, although in other clinical settings common strains in patients with cystic fibrosis are genetically indistinguishable from common environmental strains^{106,108-110}. Prevention of cross-infection has become a key part of the management of all patients and implementation of intensive infection control measures has reduced rates of shared strain infection in treatment centres^{111,112}. However, such control measures remain challenging in resource-limited settings, especially with the rapid growth in the numbers of adults with cystic fibrosis.

Non-tuberculous mycobacteria cause infection in 3–20% of patients with cystic fibrosis¹¹³, and prevalence rates have increased over the past 10 years¹¹⁴. Rapidly growing species (for example, *Mycobacterium abscessus*) are difficult to treat because prolonged courses of multiple antibiotics are required, which are often associated with toxicity and are a contraindication for lung transplantation in many transplant centres⁶⁵. Of recent concern are the reports suggesting the potential for person-to-person transmission of *M. abscessus*¹¹⁵.

Over the past several years, the airway has been revealed to not be sterile in health; the airway in cystic fibrosis is now considered to harbour a polymicrobial milieu. Both non-culture-based and culture-based methods have shown a wide range of atypical bacterial species, including *Prevotella*, *Fusobacterium* and *Veillonella* spp. among others^{116,117}. The clinical significance of the microbiome in disease progression remains to be established, but several features are emerging, including a heterogeneous microbiome composition in patient populations and lower bacterial diversity with increasing age and poorer lung function¹¹⁸.

The lung in cystic fibrosis is a microaerophilic environment and *P. aeruginosa* survives — and in fact thrives — within low oxygen tension biofilms¹¹⁹. Chronic *P. aeruginosa* infection is also associated with ongoing microevolution, which alters virulence, regulatory networks and acquisition of antimicrobial resistance

mechanisms^{120,121}. These factors probably contribute to the persistence of *P. aeruginosa*. The interaction of the host with pathogens is crucial for the development and the progression of pulmonary disease in cystic fibrosis, which leads to perpetual neutrophil recruitment to the lungs (where neutrophils are the dominant inflammatory cells)¹²¹. A key component of this dysregulation is the development of neutrophil extracellular traps (NETs), which are stimulated by bacterial pathogens (such as P. aeruginosa and S. aureus)122. NETs have detrimental effects in the cystic fibrosis airway by enhancing the viscosity of airway secretions, dampening pathogen clearance and potentially enhancing biofilm development and persistence¹²². The macrophage is likely to be an important scavenger in the lungs in patients with cystic fibrosis, but the mechanisms of action are debated. Some studies have suggested that T cells accumulate within the subepithelial airway but are limited in the lumen¹²³. Whether specific lymphocyte subsets have an important role in lung disease remains controversial. Evidence would suggest that T helper 17 $(T_{\mu}17)$ cells are important drivers of neutrophilic inflammation and an abnormal regulatory T cell response has been described to be linked to CFTR deficiency^{124,125}. The effect of polymicrobial infection is likely to add to the complexity of regulation of immune responses in the cystic fibrosis airway.

Diagnosis, screening and prevention Principles of diagnosis

The conventional diagnostic criteria for cystic fibrosis are given in BOX 2. Diagnosis is increasingly made by newborn screening (see below). However, physicians should be aware of symptoms and signs of disease in older children and adults, particularly bronchiectasis, recurrent respiratory tract infection, nasal polyps, male infertility and portal hypertension; late-presenting patients are usually — but not invariably — pancreatic sufficient. Screening can miss some mild cases if the child is born in a region without routine screening, the child missed screening despite being born in a screening region or a laboratory error occurred.

More than 95% of patients presenting symptomatically can be diagnosed with a sweat test. The test measures chloride concentration after pilocarpine (a muscarinic receptor agonist) application, which stimulates sweat production, but in a small minority of patients the diagnosis is unclear even after extensive testing¹²⁶. The upper limit of normal sweat chloride concentration remains under contention; in Europe, the upper limit of normal for an indeterminate result is 30 mmol l⁻¹ regardless of age. Given that sweat chloride concentration can increase with age, 40 mmol l⁻¹ is considered the upper limit after 6 months of age only in the United States. Above 60 mmol l⁻¹ is definitely abnormal for all ages and, provided the clinical presentation is compatible, is diagnostic¹²⁷. However, rare but unequivocal cases of cystic fibrosis have been reported with sweat chloride of <30 mmol l⁻¹, which probably occurred owing to heterogeneity in end-organ CFTR expression or the influence of other related genes such as the gene that encodes ENaC¹²⁸.



Figure 4 | **Diagnosing cystic fibrosis: nasal potential difference measurement.** Representative nasal potential difference tracings from a healthy control (blue) and a patient with cystic fibrosis (red) are shown. The nasal mucosa is sequentially perfused with Ringer's solution (an isotonic solution relative to the bodily fluids), Ringer's solution with amiloride to block the epithelial sodium channel (ENaC), chloride-free solution with amiloride, chloride-free solution with amiloride and isoproterenol to activate the cystic fibrosis transmembrane conductance regulator (CFTR), and finally the addition of ATP to activate non-CFTR-dependent anion transport. The change in potential difference upon addition of amiloride is used to estimate sodium transport, which is elevated in cystic fibrosis. The change in potential difference with chloride-free isoproterenol is used to measure CFTR-dependent anion transport, which is reduced in cystic fibrosis. Patients with mild phenotypes of cystic fibrosis generally show intermediate results.

Genetic testing is an important part of diagnostics because mutation-specific therapy is an increasing reality and can help to resolve unclear cases. Given the marked variation in the prevalence of *CFTR* mutations between different ethnic groups, test panels that account for this variation should be used unless whole-exome sequencing is carried out. In addition, the unknown significance of most of the rare *CFTR* mutations does not support the use of extended genotyping in patients with equivocal diagnostic tests¹²⁶.

Measurement of the transepithelial potential difference is an adjunct diagnostic test but is only available in a few specialist centres. The in vivo technique usually measures the potential difference across the nose or the respiratory epithelium via a catheter referenced to a peripheral electrode¹²⁹⁻¹³¹. Rectal potential difference can also be measured in vivo; alternatively, in vitro measurement of CFTR activity on excised rectal biopsy tissue can be carried out via open-circuit or closedcircuit currents^{132,133}. Other in vitro methods for assessing CFTR function, such as intestinal organoid swelling or the function of nasal epithelial cells grown in culture, are also of emerging interest¹³⁴. The nasal potential difference test in patients with cystic fibrosis has several cardinal features, including elevated potential difference at baseline, a heightened response to amiloride and a diminished response to chloride-free isoproterenol compared with healthy controls (FIG. 4). A modified sweat test that assesses β -adrenergic sweat secretion has recently been developed and might have advantages in individuals with mild disease. However, its diagnostic use is not yet well established¹³⁵. Other supportive tests can be considered to clarify the diagnosis, including stool human faecal elastase measurement for pancreatic insufficiency and, in postpubertal men, semen analysis to test for azoospermia³².

Diagnosis is hindered in patients with positive diagnostic tests but without symptoms and in patients with a cystic fibrosis phenotype but negative or equivocal diagnostic tests¹³⁶. The case for the latter patients is relatively straightforward; irrespective of the underlying diagnosis, any organ disease should be treated on its merits and the patient should be carefully monitored. Seemingly symptom-free patients with positive diagnostic tests should be carefully followed to detect the development of complications¹³⁷ whilst not overburdening the patients with treatment.

Newborn screening

Newborn screening for cystic fibrosis has been controversial because of its cost, the creation of anxiety around the procedure in seemingly healthy infants and the lack of established pulmonary treatments for infants. The cost of treatment has been shown to be reduced in patients who have been diagnosed through newborn screening compared with those who have had later diagnoses138. A randomized controlled trial has clearly shown the efficacy of screening and has provided evidence of nutritional — but not respiratory — benefits¹³⁹. However, a factor in the unexpectedly poor respiratory outcomes in that study might have been failure to apply modern infection control precautions in one of the participating centres. Much evidence shows benefit when comparing outcomes before and after the introduction of screening¹⁴⁰⁻¹⁴². For example, The London Cystic Fibrosis Collaboration showed that infants diagnosed later (in the first 2 years of life) had airway obstruction at presentation, even if they had had no respiratory symptoms or signs, and this never recovered despite specialist treatment^{143,144}. By comparison, the outcomes in babies diagnosed by screening were much more favourable (see below). Another benefit of newborn screening is that parents have the opportunity to make informed choices about antenatal diagnosis in future pregnancies.

A large number of screening protocols are available, including measurements of serum immunoreactive trypsin and CFTR mutation analysis¹⁴⁵⁻¹⁴⁷. Screening diagnosis must always be confirmed with a sweat test, not least to ensure that there has not been an error in the screening laboratory. The possible outcomes are: a definite diagnosis of classic cystic fibrosis depending on the protocol, cystic fibrosis definitely excluded or an indeterminate outcome. Indeterminate outcomes are the most challenging situations; these children fall into two groups, namely those harbouring CFTR mutations who will develop late-onset, mild-variant disease and those with true diagnostic uncertainty¹⁴⁸. Thus, screening can lead to the child being given a diagnosis of cystic fibrosis that might not have been possible until middle age. Such a situation raises the question of whether an early diagnosis affects the quality of life of the individual and their family. If accurate information is carefully given, and the intensity of treatment and monitoring is appropriately applied according to the severity of the clinical state,

quality of life need not be negatively affected. However, there is disagreement even between experts about some variants, for example, R117H with the 7T intron 8 variant in *trans*, with some believing this to be a benign variant, whereas others have identified late-presenting patients with this mutation^{149–152}.

True diagnostic uncertainty is another outcome of screening. The US Cystic Fibrosis Foundation has coined the phrase 'CFTR-related metabolic syndrome' to describe infants with sweat chloride values of <60 mmol l⁻¹ (which could be too high an upper limit of normal in infancy, given that virtually all healthy babies have a sweat chloride of <30 mmol l⁻¹) and two CFTR mutations, one of which has not been shown to be disease causing¹⁵³. An alternative term — cystic fibrosis screen positive with inconclusive diagnosis (CFSPID) has been proposed by the European Cystic Fibrosis Working Group¹⁵⁴. However, some of these infants develop cystic fibrosis-like symptoms and current recommendations are for careful follow-up monitoring¹⁵⁵. Nonetheless, despite these issues, the case in favour of newborn screening is overwhelming.

Detection of early lung disease

Infants who are diagnosed through screening are often asymptomatic with few if any clinical signs of lung disease. When investigating these infants, it is important not to miss important warning signs or to cause anxiety by overinterpreting trivial abnormalities. Current techniques to identify lung disease are lung function measurements, imaging (mainly high-resolution CT, although MRI is being increasingly used¹⁵⁶) and bronchalveolar lavage.

In terms of lung function tests, spirometry measures lung volume and air flow but is insensitive to distal airway disease. In preschool and school-age children, multi-breath washout (the lung clearance index (LCI)) is the most sensitive test in clinical practice and outperforms spirometry and lung volume assessment by body plethysmography¹⁵⁷⁻¹⁵⁹. However, in the first 2 years of life, LCI will fail to detect a substantial number of children who have abnormal infant spirometry, so the two tests should be combined in this age group¹⁶⁰. Two large collaborative projects have evaluated lung function in infants, with differing results: although both groups reported that lung function was abnormal at diagnosis^{161,162}, the Australian Respiratory Early Surveillance Team for Cystic Fibrosis (AREST-CF) reported rapid and marked deterioration¹⁶³, whereas the London Cystic Fibrosis Collaboration (LCFC) showed improvement over the first year of life, with stabilization in the second year^{164,165}. These differences could be explained by differences in treatment strategies or in the characteristics of the population studied.

LCI has also been used as a clinical trial end point; one study has shown that an elevation in LCI predicts pulmonary exacerbations¹⁶⁶⁻¹⁶⁸. The role of highresolution CT is controversial owing to the radiation exposure and the potentially increased baseline risk of some epithelial cancers in patients with cystic fibrosis¹⁶⁹. Indeed, its role in infants diagnosed through screening is debated; the AREST-CF group has shown a large number of lung abnormalities⁸⁹, whereas the LCFC group found that CT changes at 1 year of age were so mild that they could not be reproducibly scored, and accordingly abandoned the technique¹⁷⁰. In schoolage children, if LCI is abnormal then the CT will also probably be abnormal and the number of scans could, therefore, be reduced^{171,172}. If high-resolution CT is carried out, dilated airways should not be assumed to be 'bronchiectatic' (that is, irreversibly dilated) as airtrapping might simply reflect transient mucus plugging rather than any structural disease. No study has shown that regular CT scans improve prognosis, and scanning should be reserved for selected cases.

Identification of lower respiratory tract pathogens can be hampered by the patients' inability to spontaneously produce a sputum sample; for these patients, bronchoalveolar lavage can be considered. Early bronchoalveolar lavage surveillance programmes have shown that infection and inflammation in the absence of symptoms is common^{88,98,173}. A 5-year, prospective randomized controlled trial showed no benefit from an aggressive programme of regular bronchoscopies; this technique should only be reserved for children who are symptomatic despite antibiotic treatment¹⁷⁴. Early elevation of bronchoalveolar lavage neutrophil elastase activity has been shown to predict later structural changes on CT⁸⁹, but no evidence currently supports that detecting neutrophil elastase facilitates an intervention that improves prognosis.

Extrapulmonary manifestations

Cystic fibrosis is a multisystem disease that affects many organs in which CFTR is expressed. Chronic rhinosinusitis is extremely common and nasal polyposis a complication in up to 45% of patients^{175,176}. Persistent infection in the upper airways and sinuses can be a source of lower respiratory tract infection¹⁷⁷. The gastrointestinal tract is affected in numerous ways, including increased nutrient loss secondary to pancreatic insufficiency, reduced fat-soluble vitamin absorption, fat-soluble vitamin deficiency states, frequent gastro-oesophageal reflux and impaired bowel transit (complicated by distal intestinal obstruction syndrome, constipation and small intestinal bacterial overgrowth)¹⁷⁸⁻¹⁸². The early recognition of nutritional deficits is vital, as poor growth and malnutrition adversely affect pulmonary function and patient survival183,184. Hepatic involvement is also common, with up to one in three patients affected including those with evidence of hepatic steatosis, cholelithasis, ductal stenosis and focal biliary cirrhosis^{185,186}. Biliary cirrhosis usually becomes clinically evident in late childhood or early adolescence and leads to portal hypertension.

Cystic fibrosis-related diabetes is increasingly common with advancing age and is clinically distinct from type 1 and type 2 diabetes mellitus in the general population¹⁸⁷. Importantly, its occurrence adversely affects survival in patients with cystic fibrosis¹⁸⁸. Other endocrinological complications of cystic fibrosis include delayed menarche in malnourished adolescent females and reduced bone mineral density,

which can increase the risk of bone fractures and is multifactorial¹⁸⁹. Congenital bilateral absence of the vas deferens occurs in 98% of males with cystic fibrosis and results in azoospermia; intracytoplasmatic sperm injection or related procedures are required for these men to father children. Renal complications can occur and include nephrocalcinosis, and salt and water depletion owing to excess fluid losses, which contribute to acute kidney injury and proteinuria. These renal complications can occur even in those without evidence of diabetes. Chronic kidney disease is more common in adult patients and risk factors for its development include age, diabetes, prior episodes of acute kidney injury and prior organ transplantation. As survival for patients with cystic fibrosis has increased, several complications have been recognized, and these include an increased risk of colorectal and other gastrointestinal malignancies, the potential for macrovascular disease to complicate longstanding inflammatory disease and venous insufficiency.

Management

Clinical outcomes in cystic fibrosis vary between countries and centres^{56,190-193}, and evidence for many areas of clinical decision making is lacking. Thus, the current focus is on benchmarking projects⁵⁶, the establishment of standards of care and management guidelines^{194,195} and the development of multidisciplinary specialist care programmes that work in collaboration with the patient and their family¹⁹⁶. National and international data registries of patients with cystic fibrosis were established in the 1960s^{18,197,198}. These registries are central for clinical research and care^{15,19,55,199,200}. Public reporting of data and benchmarking between centres have led to improvements of clinical outcomes in less well-achieving centres and high-achieving centres^{56,201}.

Over the past 20 years, therapies and the time and effort required by patients and their families have markedly increased. Not surprisingly, adherence to prescribed therapies is quite low²⁰², with lowest rates of adherence observed for airway clearance and nebulized medications. One US study indicated that, overall, adherence to pulmonary therapies is \leq 50%, with decreases in adherence related to increasing age from childhood to adulthood^{203,204}. Similarly, use of complex and life-long therapies has led to the emergence of treatment-related toxicities, β -lactam allergies, aminoglycoside nephrotoxicity and vascular complications from long-term intravenous access devices²⁰⁵.

Early recognition and treatment of lung disease²⁰⁶ and exacerbations^{207,208}, *P. aeruginosa* eradication and prevention of chronic infection, optimization of nutritional status, adequate mobilization of airway secretions, physical fitness and psychosocial management require regular consideration²⁰⁸. Indeed, with the adult cystic fibrosis population growing rapidly, comprehensive transitional care programmes should be available at all centres with regular communication between paediatric and adult teams^{208,209}. The key components of pulmonary care of all people with cystic fibrosis are knowledge, participation and monitoring of airway clearance and the appropriate use of maintenance therapies such as mucolytic therapy, hydrators and antibiotics (FIG. 5).

Pulmonary disease

Airway clearance and exercise. Daily airway clearance is considered a standard of care, although the strength of evidence to support long-term benefit is limited²¹⁰. Modes of airway clearance include percussion, device assistance (for example, positive expiratory pressure and vest and handheld vibratory devices) and breathing modalities (for example, autogenic drainage). Although few comparative trials have been carried out, one has shown positive expiratory pressure to be associated with a lower rate of pulmonary exacerbations — an outcome measure that is associated with lung function decline — compared with an oscillating vest device²¹¹. Aerobic exercise results in improved exercise tolerance, and increased physical activity has been linked to reduced lung function decline^{212,213}.

Mucolytic and hydrator therapies. Dornase alfa, which is a recombinant human deoxyribonuclease, breaks down DNA derived from degrading neutrophils that accumulate in the airways of patients with cystic fibrosis, thereby reducing viscosity of airway secretions and leading to improved lung function and reduced exacerbations^{214,215}. The first large-scale study to show benefit of nebulized dornase alfa was reported >20 years ago²¹⁴. Benefits have subsequently been shown in patients with advanced lung disease (FEV₁ <40% of predicted) and in younger patients with mild disease^{216,217}. This therapy is considered the standard of care in patients ≥5 years of age^{208,218}.

Hypertonic saline functions as a hydrating agent that increases mucociliary clearance and has been shown to improve lung function and to reduce exacerbations in a randomized controlled trial²¹⁹. A reduction in exacerbations was not observed in a study of children aged 4 months to 5 years, but substudies have shown positive effects on lung function^{220,221}. Additional studies to assess the efficacy of hypertonic saline in younger children are currently ongoing. Inhaled dry powder mannitol is another osmotic agent that has shown lung function improvements in two trials; effects have been more consistent in adults than in children^{222,223}.

Inhaled antibiotics. In patients with chronic P. aeruginosa infection, nebulized tobramycin (an aminoglycoside antibiotic) improved lung function, reduced exacerbations and increased weight^{224,225}. A dry powder preparation of tobramycin that decreases delivery time has more recently been shown to have equal efficacy and to improve patient satisfaction²²⁶. Inhaled aztreonam (a β -lactam antibiotic) has also been shown to be efficacious when compared with both placebo and inhaled tobramycin^{227,228}. Other antibiotics such as colistin are used in some countries and several new preparations of inhaled antibiotics are currently being studied²²⁹. Few studies have so far examined the role of inhaled antibiotics for other bacterial infections that are common in patients with cystic fibrosis (for example, B. cepacia complex, MRSA and S. Maltophilia)²³⁰.



Figure 5 | **Currently available therapies to treat patients with cystic fibrosis.** Cystic fibrosis transmembrane conductance regulator (CFTR) therapies are based on the cellular mechanism, generally address a specific aspect of the molecular defect and do not therefore alleviate the effects of all classes of CFTR mutations. The CFTR potentiator ivacaftor directly activates CFTR and is currently licensed in most countries for patients with class III (gating) mutations. Hypertonic saline increases airway surface liquid, which is reduced in patients with cystic fibrosis as a consequence of defective chloride and increased sodium absorption. Dornase alfa cleaves extracellular DNA, thereby reducing the viscosity of airway secretions. Inhaled tobramycin and aztreonam are used as chronic maintenance therapy in patients with chronic *Pseudomonas aeruginosa* infection. Azithromycin has multiple potential modes of action but mainly ameliorates airway inflammation, as is the case for high-dose ibuprofen, which reduces neutrophil influx into the airways.

Macrolides. Several randomized controlled trials have shown improved lung function, quality of life, weight and reduced time to next exacerbations in *P. aeruginosa*-infected patients treated with azithromycin^{231,232}. The mechanism of action of macrolides in cystic fibrosis might be anti-inflammatory rather than antibacterial. In patients without *P. aeruginosa*, when treated with azithromycin, lung function improvement was not observed but there was evidence of a reduced

pulmonary exacerbation rate^{233,234}. An increasing prevalence of nontuberculous mycobacteria in a singlecentre study has raised concerns of whether long-term azithromycin treatment is a contributory factor. This finding was not supported by a subsequent national data registry analysis^{113,115}, but the effect of macrolides on nontuberculous mycobacterial infection remains controversial as complete case ascertainment is unlikely in registry analysis.

Anti-inflammatory therapies. Several anti-inflammatory therapies have been studied in patients with cystic fibrosis, including oral corticosteroids, inhaled corticosteroids, non-steroidal anti-inflammatory drugs (such as ibuprofen) and leukotriene inhibitors. Although ibuprofen reduced the rate of decline in lung function, especially in younger patients and adolescents, its uptake has been limited because of the need to monitor blood levels to optimize benefit^{235,236}. Oral steroids are associated with adverse side effects and inhaled steroids have limited effect in patients who do not have asthma in addition to cystic fibrosis²³⁷⁻²⁴⁰. A leukotriene B4 inhibitor (known as BIIL 284) was associated with increased exacerbations and led to an early termination of one trial, raising concerns that anti-inflammatory therapies involve a delicate balance in reducing inflammation without adversely affecting the patient's response to infection²⁴¹.

Lung transplantation. Lung transplantation is the established treatment for patients with end-stage pulmonary disease. The outcomes for patients undergoing transplantation for cystic fibrosis have rapidly improved and median survival now approaches or even exceeds 10 years in many treatment centres. Timely referral and close communication between the cystic fibrosis and transplant centres is required to provide sufficient time for assessment of suitability (of the donor and of the recipient), to determine whether indications are met and to establish that there are no foreseeable contraindications. Donor allocation programmes vary globally but aim to prioritize those waiting for transplants who have the most limited pre-transplant survival²⁴². Adjunctive therapies, including noninvasive ventilation, can function as a bridge to transplantation²⁴³. The most common cause of graft failure following lung transplantation is bronchiolitis obliterans, which is thought to be a form of chronic allograft rejection²⁴².

Extrapulmonary disease

As median survival from cystic fibrosis has approached or even exceeded 40 years, complications have emerged. For example, gastrointestinal malignancy, hyperlipidaemia, metabolic and endocrine complications and multiresistant infections have emerged and are providing new challenges²⁰⁵. Involvement of an interdisciplinary team and access to specialist support, including experts in microbiology, general surgery, thoracic surgery, gastroenterology and hepatology, otolaryngology, obstetrics and gynaecology, clinical genetics, endocrinology, palliative care and transplantation services are increasingly important for the adult with cystic fibrosis²⁰⁸.

Treatment of the basic defect

In terms of treating the basic genetic defect, therapies targeting CFTR dysfunction work by inserting a normal copy of *CFTR* into patients cells (gene therapy), improving the expression of CFTR on the cell surface, increasing the 'opening probability' of existing channels (CFTR pharmacotherapy) or by targeting other ion channels to compensate for its dysfunction.

Gene therapy. Somatic gene therapy has mainly been studied as topical therapy administered to the airways to minimize systemic toxicity. The technique enables transient expression of mature and functional CFTR on the cell surface, as shown in cell cultures and in mice²⁴⁴. Human trials initially focused on adenoviral vectors owing to their high transfection efficiency *in vitro*. Single-dose trials involving nasal or intra-tracheal administration showed transient expression of CFTR without substantial adverse effects at low doses^{245,246}. However, repeated dosing is associated with an immune response that reduces transfection efficiency and clinical benefit has not yet been shown²⁴⁷. Modifications of adenoviral vectors might help to reduce their immunogenic potential.

Using adeno-associated virus as the vector is less immunogenic than adenoviral vectors and initial studies have suggested prolonged expression and positive trends in clinical outcome measures²⁴⁸. However, this finding was not subsequently supported in a longer term multidose trial²⁴⁹. Lentiviral vectors are currently being investigated in preclinical studies, in which they have shown high transfection efficiency and prolonged activity of CFTR expression²⁵⁰.

Liposomal vectors could potentially overcome these limitations of viral vectors. Liposomes are less immunogenic and therefore better suited for repeated dosing; unfortunately, this positive attribute is associated with a lower level of transfection efficiency. A multicentre multiple dose trial has recently been completed by The UK Cystic Fibrosis Gene Therapy Consortium and results are expected soon²⁵¹. Delivery of RNA rather than DNA is another strategy that is currently being explored²⁵².

Translational readthrough therapy. Class I mutations include premature termination codons (PTCs) that lead to a truncated protein and mRNA transcripts that undergo nonsense-mediated decay. Aminoglycosides, such as gentamicin, can bind to the ribosome and enable readthrough of PTCs, which leads to some production of the full-length protein and partial restoration of channel function. Both *in vitro* studies and studies using topical application to the nasal epithelium in patients carrying stop mutations on at least one allele have shown evidence of CFTR expression after gentamicin treatment²⁵³. Clinical applicability is limited as prolonged administration of gentamicin is required, which would probably cause renal and/or ototoxicity.

Ataluren was developed as a compound with similar translational readthrough properties but it lacks the potential adverse effects of aminoglycosides²⁵⁴. Initial uncontrolled studies in patients showed improvement in chloride conductance measured by nasal potential difference, but a larger placebo controlled trial failed to show clinical benefit^{19,255}. However, the subgroup of patients not receiving inhaled antibiotics and without chronic exposure showed less lung function decline after 12 months, an effect that could be explained by competitive inhibition at the level of the ribosome. One study²⁵⁶ is under way to prospectively assess efficacy in this subgroup. Overall, the benefits observed so far are

Box 3 | Domains on the Cystic Fibrosis Questionnaire-Revised

Functional and psychosocial scales

- Physical functioning: ability to walk, climb stairs and carry heavy items, and ability to run, jump and play
- Social and school functioning: going out with friends and engaging in social activities
- Emotional functioning: feeling happy, sad or worried
- Treatment burden: time spent on treatments and fitting treatments into daily activities
- Eating problems: challenges eating and making calorie goals
- Body image: physical appearance and being short or thin
- Vitality*: energy level and extent of fatigue
- Health perceptions*: perceptions of current health and disease severity
- Role functioning*: ability to carry out daily activities (attending school, working and household tasks)

Symptom scales

- Respiratory symptoms: frequency and severity of cough, mucus production and chest congestion
- Digestive symptoms: frequency and severity of abdominal pain, stools and gas
- Sinus symptoms: frequency and severity of sinus headaches, nasal congestion and postnasal drip
- Weight*: perception of current weight

*Not administered to children 6–13 years of age

limited in magnitude, but could probably be increased by combining ataluren with a CFTR potentiator.

CFTR potentiator therapy. Mutations that cause residual surface expression are potentially amenable to drugs that increase channel opening probability. These drugs are called potentiations and the first drug, ivacaftor, has been approved in most countries for clinical use in patients with class III mutations. The most common CFTR class III mutation, G551D, is associated with normal cell surface expression but reduced gating. Ivacaftor improves CFTR function, as shown by improvement in ion channel measurements^{24,25,257}. Notably, sweat chloride concentrations fell below the diagnostic threshold in most treated patients - a result that has been confirmed in an observational study²⁵⁸. This drop in sweat chloride is accompanied by marked improvement in lung function as well as improvements in other clinical measures (weight, symptoms and pulmonary exacerbations)257. Similar effects have been described in other gating mutations, which underlines the potential of CFTR pharmacotherapy in a broader range of CFTR mutations^{258,259}.

Studies are also underway in other mutation classes that could benefit from potentiation of CFTR function, including class IV and class V mutations, as well as mutations in class VI that are associated with residual CFTR function. In addition, as readthrough and corrector therapy is unlikely to normalize CFTR function alone, potentiators might be needed for combination therapy in patients carrying class I or class II mutations. Potentiation of CFTR function could also be of benefit in diseases with secondary CFTR dysfunction, such as chronic bronchitis, as smoke exposure has been found to cause secondary CFTR dysfunction²⁶⁰.

Intracellular trafficking. The most common CFTR mutation, F508del, is associated with defective protein folding that results in proteosomic degradation with very little or no CFTR reaching the apical membrane (class II). Indeed, as little or no CFTR is expressed on the cell surface in F508del homozygous patients, potentiators such as ivacaftor have little effect on chloride transport in bronchial epithelial cells; this finding has recently been confirmed in a Phase II study²⁶¹. Drugs affecting CFTR transport have been called 'correctors' even though they do not necessarily correct the folding defect. Some correctors alter cell chaperones and other quality control mechanisms; nevertheless, their use generally increases trafficking to the cell surface. The best-studied compounds have evolved from highthroughput screening programmes, with two small molecules — lumacaftor and VX-661 — having undergone clinical studies. Given alone, both compounds have little clinical benefit in patients with F508del; in fact, lung function worsened in patients on lumacaftor alone²⁶². However, when ivacaftor was added to lumacaftor, lung function improved above baseline²⁶². The magnitude of the effect was significantly less than that observed in response to ivacaftor alone in patients harbouring the G551D mutation (class III), which corroborates results from in vitro experiments in human bronchial epithelial cells²⁶³. Recent in vitro data suggest that ivacaftormediated CFTR activation might destabilize F508del CFTR and might decrease the beneficial effects of some correctors, including lumacaftor; whether this finding is relevant in vivo is currently unclear, but this interaction could be one explanation for the fairly small clinical effect of combination therapy with ivacaftor and lumacaftor in F508del homozygous patients^{264,265}. Although the clinical efficacy data of combination therapy is promising, it has become clear that the search for additional CFTR corrector compounds needs to continue. Adding a second corrector has been shown to improve the efficacy of lumacaftor and ivacaftor in vitro266 by addressing distinct cellular mechanisms.

Ion channel-directed therapy. Cystic fibrosis is associated with decreased chloride and bicarbonate secretion through CFTR and increased sodium absorption by ENaC. Another potential mechanism to address the ion dysbalance on the epithelial surface is to target other apical ion channels. Activation of the apical calcium-activated chloride channel using derivatives of its natural activator ATP has been studied using the P2Y purinoceptor agonist denufosol. Although initial studies showed some promise, beneficial effects could not be confirmed in subsequent studies^{267,268}. Inhibition of sodium absorption was initially examined using amiloride, but limited clinical efficacy was shown²⁶⁹. More potent and specific inhibitors with longer half-lives are currently being developed.

Quality of life

There is growing recognition of the importance of patient-reported outcomes (PROs) in health outcomes research and clinical care²⁷⁰. These instruments, which



Figure 6 | **Emerging approaches to address the ion channel abnormalities in cystic fibrosis.** The main target for therapeutics is cystic fibrosis transmembrane conductance regulator (CFTR), which can be replaced through gene therapy or increased in concentration on the cell surface. Increased CFTR concentration could be achieved through translational readthrough therapy (class I mutations), correction of intracellular trafficking (class II mutations) or potentiation of its function (class III mutations and potentially class IV and class V). Alternatively, a calcium-activated chloride channel could be a therapeutic target; this channel is activated through the P2Y purinoceptor 2 (P2Y2; the natural ligand being ATP). Finally, as sodium absorption is upregulated in the airways of patients with cystic fibrosis, blocking the epithelial sodium channel (ENaC) could be of benefit to increase airway surface liquid.

include health-related quality-of-life (HRQOL) measures, can be used for several purposes: as primary or secondary outcomes in clinical trials of new medications or behavioural interventions^{219,271,272}; to document the natural progression of the disease²⁷³; to describe the effect of an illness on patient functioning²⁷⁴; to analyse the costs and the benefits of medical interventions; and to aid in communication and clinical decision making²⁷⁵.

Quality-of-life studies in cystic fibrosis over the past 10 years have highlighted three key issues: the importance of patient-reported respiratory symptoms as an outcome measure for clinical trials; the growing perceptions among patients and families that the prescribed treatment regimen is burdensome²⁷⁶; and the considerable differences in HRQOL according to socioeconomic and racial and/or ethnic minority status. Indeed, prior research on HRQOL in cystic fibrosis has shown that generic measures are insensitive and that disease-specific tools are needed²⁷⁷. Efforts to develop reliable and valid HRQOL instruments in cystic fibrosis have been successful^{228,270,278} and are now routinely used in clinical trials of new medications and in studies of patient functioning. Currently, the most widely used HRQOL measure for cystic fibrosis is the Cystic Fibrosis Questionnaire-Revised (CFQ-R) (BOX 3), with developmentally appropriate versions for preschooler children, school-aged children and their parents, and adolescents and adults²⁷⁹. The CFQ-R has been translated into more than 36 languages and is being used in several multinational trials.

Randomized controlled trials of new therapies in cystic fibrosis have used the CFQ-R as a primary or secondary outcome and have shown benefits in terms of reduction in respiratory symptoms and improvements in other domains, such as physical functioning, vitality and health perceptions^{228,271,273,279}. Interestingly, improvements in respiratory symptoms have been found across trials of medications with very different mechanisms of actions (for example, inhaled antibiotics, hypertonic saline and potentiators), increasing confidence that this is an important outcome.

Although a primary reason that health outcomes and lifespan have markedly improved over the past 20 years has been the development of new long-term therapies for patients with cystic fibrosis, treatment regimens now take 2–3 hours per day for most patients²⁷⁹. In several studies using the CFQ-R, which has a treatment burden scale, patients and parents have reported increasing perceptions of burden^{274,280}. In a recent US epidemiological study over 3 years, both treatment complexity and perceived burden were highest among adults and those with severe lung disease, but increased in all age groups over the course of the study²⁸¹.

Using a national database in the United States, differences on the CFQ-R were shown by socioeconomic and racial and/or ethnic minority status. In a sample of 4,751 patients and 1,826 parents, after controlling for disease severity, people with low socioeconomic status reported significantly lower scores on the CFQ-R (children, parents and teen/adults) across the majority of domains. After controlling for both disease severity and socioeconomic status, African-American and Hispanic families reported lower scores on the social and emotional functioning scales compared with their white counterparts²⁸². These differences might be related to access to care, ability to afford medical insurance and other ancillary costs of optimal care (for example, good nutrition). Whether similar disparities are found in countries with national healthcare systems remains unclear.

Another important issue that affects HRQOL is comorbid depression and anxiety. An international psychological screening study of >6,000 patients with cystic fibrosis and 4,000 parent caregivers in nine countries recently found a significantly higher prevalence of these symptoms than in community samples²⁸³. Psychological distress has a direct negative effect on therapy adherence, attendance at clinic, hospitalizations and healthcare costs^{284–287}. As we move towards more patient-centred care, standardized assessments of HRQOL and psychological symptoms will be increasingly integrated into clinical care.

Outlook

Understanding the underlying genetic abnormalities and the mechanisms by which CFTR mutations cause disease has led to CFTR-specific therapies that are either already available to patients or that are close to clinical use (FIG. 6). Additional compounds are being assessed in ongoing studies; thus, the armamentarium of treatment options will probably expand. Although genotype-specific therapy is being referred to as personalized medicine, it will not provide the same benefit to each patient. Some studies have already shown that response to interventions is not homogeneous even in patients carrying the same CFTR mutation²⁶². Exploring modifier genes of involved intracellular pathways or other ion channels might help to better understand this variability and to define additional therapeutic targets. In addition, current studies focus on patients with the CFTR mutations with known functional consequences on CFTR processing and function, and will not be applicable to patients with rare mutations.

An individualized treatment approach will probably be needed for CFTR-directed pharmacotherapy, particularly for patients with rare or difficult to treat alleles. Individualized treatment will require test systems or biomarkers that have a high precision-predicting treatment response. Potential assays could involve cells harvested from patients that can be used to test the most promising treatment combination *in vitro*. On the basis of their accessibility, nasal epithelial cells and intestinal cells transformed into organoids are currently being investigated as potential models¹³⁴. Preliminary data using stem cells derived from skin fibroblasts or blood cells transformed into airway epithelial cells could offer an alternative in the near future²⁸⁸. Alternatives or additions to these *in vitro* assays could be used topically *in vivo* to test drug response by applying them to the sweat gland or to the nasal epithelium. In addition, sensitive clinical outcome measures will need to be developed to enable individual patient monitoring of drug response.

CFTR-directed therapy should ultimately prevent lung damage; to do so will require treatment early in the disease process, ideally as soon as the diagnosis is established. Newborn screening offers a unique opportunity, but further work is needed to establish robust outcome measures in young children to provide evidence for efficacy of early treatment interventions. Prevention of deterioration rather than improvements in disease manifestations will probably need to be the outcome measure of clinical trials in this age group. If this can be achieved, CFTR modulation could potentially maintain lung health and preserve lung function and structure. In addition, early application of CFTR modulators could also affect other disease manifestations, such as exocrine and endocrine pancreatic function. Conversely, in patients with established disease, the best scenario might be to transform cystic fibrosis into non-cystic-fibrosis bronchiectasis — a disease that is much less severe but often still progressive. Thus, even with optimal CFTR-directed treatment, it will take decades until we can expect patients to not need treatment for the prominent disease manifestations of mucus obstruction, infection and inflammation. Although targeting CFTR is likely to make a major difference, finding better therapeutic strategies for these aspects of the disease will continue to be important to maintain lung health in patients with cystic fibrosis.

- Weiler, C. A. & Drumm, M. L. Genetic influences on cystic fibrosis lung disease severity. *Front. Pharmacol.* 4, 40 (2013).
- Jarvi, K. *et al.* Cystic fibrosis transmembrane conductance regulator and obstructive azoospermia *Lancet* 345, 1578 (1995).
- Gonska, T. *et al*. Role of cystic fibrosis transmembrane conductance regulator in patients with chronic sinopulmonary disease. *Chest* 142, 996–1004 (2012).
- Wilschanski, M. *et al.* Mutations in the cystic fibrosis transmembrane regulator gene and *in vivo* transepithelial potentials. *Am. J. Respir. Crit. Care Med.* **174**, 787–794 (2006).
- Riordan, J. R. *et al.* Identification of the cystic fibrosis gene: cloning and characterization of complementary DNA. *Science* 245, 1066–1073 (1989).
- Rommens, J. M. *et al.* Identification of the cystic fibrosis gene: chromosome walking and jumping. *Science* 245, 1059–1065 (1989).
- Ratjen, F. & Grasemann, H. New therapies in cystic fibrosis. *Curr. Pharm. Des.* 18, 614–627 (2012).
 O'Sullivan, B. P. & Freedman, S. D. Cystic fibrosis
- O'Sullivan, B. P. & Freedman, S. D. Cystic fibrosis. Lancet 373, 1891–1904 (2009).
- Walters, S., M. A. in *Hodson and Geddes' Cystic* Fibrosis 3rd edn (eds Hodson, M. E., Geddes D. M., Bush, A.) 21–45 (Edward Arnold Ltd, 2007).
- Farrell, P. M. The prevalence of cystic fibrosis in the European Union. J. Cyst. Fibros. 7, 450–453 (2008).
- Festini, F. *et al.* Incidence of cystic fibrosis in the Albanian population. *Pediatr. Pulmonol.* 43, 1124–1129 (2008).
- Festini, F., Taccetti, G., Cioni, M. L., Repetto, T. & De Martino, M. High incidence of cystic fibrosis in children born in Italy to Albanian immigrants. *Thorax* 58, 93 (2003).
- Dupuis, A., Hamilton, D., Cole, D. E. & Corey, M. Cystic fibrosis birth rates in Canada: a decreasing trend since the onset of genetic testing. *J. Pediatr.* 147, 312–315 (2005).

- Kulich, M., Rosenfeld, M., Goss, C. H. & Wilmott, R. Improved survival among young patients with cystic fibrosis. J. Pediatr. 142, 631–636 (2003).
- MacKenzie, T. et al. Longevity of patients with cystic fibrosis in 2000 to 2010 and beyond: survival analysis of the cystic fibrosis foundation patient registry. Ann. Intern. Med. 161, 233–241 (2014).
- Cystic Fibrosis Foundation Cystic Fibrosis Foundation Patient Registry 2012 annual data report. (Cystic Fibrosis Foundation, 2013).
- Dodge, J. A., Lewis, P. A., Stanton, M. & Wilsher, J. Cystic fibrosis mortality and survival in the UK: 1947–2003. *Eur. Respir. J.* 29, 522–526 (2007). References 15 and 17 show the improvements in survival in the US and the UK cystic fibrosis populations, respectively, over the past decade.
- Cystic Fibrosis Canada. Canadian cystic fibrosis patient data registry report 2010. (Cystic Fibrosis Canada, 2010).
- Kerem, E. *et al*. Factors associated with FEV1 decline in cystic fibrosis: analysis of the ECFS patient registry. *Eur. Respir. J.* 43, 125–133 (2014).
- Rodman, D. M. *et al.* Late diagnosis defines a unique population of long-term survivors of cystic fibrosis. *Am. J. Respir. Crit. Care Med.* **171**, 621–626 (2005).
- Nick, J. A. et al. Effects of gender and age at diagnosis on disease progression in long-term survivors of cystic fibrosis. Am. J. Respir. Crit. Care Med. 182, 614–626 (2010).
- Simmonds, N. J. *et al.* Cystic fibrosis and survival to 40 years: a study of cystic fibrosis transmembrane conductance regulator function. *Eur. Respir. J.* **37**, 1076–1082 (2011).
- George, P. M. *et al.* Improved survival at low lung function in cystic fibrosis: cohort study from 1990 to 2007. *BMJ* 342, d1008 (2011).
- Accurso, F. J. *et al.* Effect of VX-770 in persons with cystic fibrosis and the C551D-CFTR mutation. *N. Engl. J. Med.* 363, 1991–2003 (2010).

- Ramsey, B. W. *et al.* A CFTR potentiator in patients with cystic fibrosis and the G551D mutation. *N. Engl. J. Med.* 365, 1663–1672 (2011). This article shows the efficacy of CFTR modulation by the potentiator ivacaftor in patients with the G551D mutation.
- Kerem, B. *et al.* Identification of the cystic fibrosis gene: genetic analysis. *Science* 245, 1073–1080 (1989).
- Higgins, C. F. ABC transporters: from microorganisms to man. Annu. Rev. Cell Biol. 8, 67–113 (1992).
- Klein, I., Sarkadi, B. & Varadi, A. An inventory of the human ABC proteins. *Biochim. Biophys. Acta* 1461, 237–262 (1999).
- Collins, F. S. Cystic fibrosis: molecular biology and therapeutic implications. *Science* 256, 774–779 (1992).
- Šerohijos, A. W. *et al.* Phenylalanine-508 mediates a cytoplasmic-membrane domain contact in the CFTR 3D structure crucial to assembly and channel function. *Proc. Natl Acad. Sci. USA* **105**, 3256–3261 (2008).
- Drumm, M. L. *et al.* Genetic modifiers of lung disease in cystic fibrosis. *N. Engl. J. Med.* **353**, 1443–1453 (2005).
- Chillon, M. et al. Mutations in the cystic fibrosis gene in patients with congenital absence of the vas deferens. N. Engl. J. Med. 332, 1475–1480 (1995).
- Sharer, N. *et al.* Mutations of the cystic fibrosis gene in patients with chronic pancreatitis. *N. Engl. J. Med.* 339, 645–652 (1998).
- Cohn, J. A. *et al.* Relation between mutations of the cystic fibrosis gene and idiopathic pancreatitis. *N. Engl. J. Med.* 339, 653–658 (1998).
- Sosnay, P. R. *et al.* Defining the disease liability of variants in the cystic fibrosis transmembrane conductance regulator gene. *Nat. Genet.* 45, 1160–1167 (2013)
- Welsh, M. J. & Smith, A. E. Molecular mechanisms of CFTR chloride channel dysfunction in cystic fibrosis. Cell 73, 1251–1254 (1993).

- Okiyoneda, T. *et al.* Peripheral protein quality control removes unfolded CFTR from the plasma membrane. *Science* **329**, 805–810 (2010).
- Mendoza, J. L. *et al.* Requirements for efficient correction of DeltaF508 CFTR revealed by analyses of evolved sequences. *Cell* **148**, 164–174 (2012).
 Rabeh W M *et al.* Correction of both NBD1
- Rabeh, W. M. *et al.* Correction of both NBD1 energetics and domain interface is required to restore ΔF508 CFTR folding and function. *Cell* **148**, 150–163 (2012).

References 38 and 39 delineate the importance of two distinct molecular defects conferred by the F508del *CFTR* mutation, including their effect of correcting *CFTR* misfolding to restore *CFTR* expression. These papers show that to fully restore mutated *CFTR* to the cell surface, both NBD1 stability and interdomain assembly must be addressed.

- Green, D. M. *et al.* Heritability of respiratory infection with *Pseudomonas aeruginosa* in cystic fibrosis. *J. Pediatr.* **161**, 290–295 e1 (2012).
- Garred, P. et al. Association of mannose-binding lectin gene heterogeneity with severity of lung disease and survival in cystic fibrosis. J. Clin. Invest. 104, 431–437 (1999).
- Sun, L. et al. Multiple apical plasma membrane constituents are associated with susceptibility to meconium ileus in individuals with cystic fibrosis. Nat. Genet. 44, 562–569 (2012).
- Blackman, S. M. *et al.* Genetic modifiers of cystic fibrosis-related diabetes. *Diabetes* 62, 3627–3635 (2013).
- Dorfman, R. *et al.* Modifier gene study of meconium ileus in cystic fibrosis: statistical considerations and gene mapping results. *Hum. Genet.* **126**, 763–778 (2009).
- Romi, H. *et al.* Meconium ileus caused by mutations in *GUCY2C*, encoding the CFTR-activating guanylate cyclase 2C. *Am. J. Hum. Genet.* **90**, 893–899 (2012).
- Li, W. et al. Unraveling the complex genetic model for cystic fibrosis: pleiotropic effects of modifier genes on early cystic fibrosis-related morbidities. Hum. Genet. 133, 151–161 (2014).
- Wright, F. A. *et al.* Genome-wide association and linkage identify modifier loci of lung disease severity in cystic fibrosis at 11p13 and 20q13.2. *Nat. Genet.* 43, 539–546 (2011).
- Collaco, J. M., Blackman, S. M., McGready, J., Naughton, K. M. & Cutting, G. R. Quantification of the relative contribution of environmental and genetic factors to variation in cystic fibrosis lung function. *J. Pediatr.* **157**, 802–807 e1–3 (2010).
- Vanscoy, L. L. *et al.* Heritability of lung disease severity in cystic fibrosis. *Am. J. Respir. Crit. Care Med.* **175**, 1036–1043 (2007).
- Kolbe, E. W. *et al. CLCA4* variants determine the manifestation of the cystic fibrosis basic defect in the intestine. *Eur. J. Hum. Genet.* **21**, 691–694 (2013).
 Bremer, L. A. *et al.* Interaction between a novel *TGFB1*
- Bremer, L. A. *et al.* Interaction between a novel *TGFB1* haplotype and *CFTR* genotype is associated with improved lung function in cystic fibrosis. *Hum. Mol. Genet.* 17, 2228–2237 (2008).
- Yarden, J. *et al.* Association of tumour necrosis factor alpha variants with the CF pulmonary phenotype. *Thorax* 60, 320–325 (2005).
- Goss, C. H., Newsom, S. A., Schildcrout, J. S., Sheppard, L. & Kaufman, J. D. Effect of ambient air pollution on pulmonary exacerbations and lung function in cystic fibrosis. *Am. J. Respir. Crit. Care Med.* 169, 816–821 (2004).
- Collaco, J. M. *et al.* Interactions between secondhand smoke and genes that affect cystic fibrosis lung disease. *JAMA* 299, 417–424 (2008).
- Martin, B. *et al.* Comparison of the US and Australian cystic fibrosis registries: the impact of newborn screening. *Pediatrics* **129**, e348–e355 (2012).
 Boyle, M. P., Sabadosa, K. A., Quinton, H. B.,
- Boyle, M. P., Sabadosa, K. A., Quinton, H. B., Marshall, B. C. & Schechter, M. S. Key findings of the US Cystic Fibrosis Foundation's clinical practice benchmarking project. *BMJ Qual. Saf.* 23 (Suppl. 1), i15–i22 (2014).

This article reports the key outcomes of the quality improvement programme of cystic fibrosis care in the United States and shows the assessment of patterns of practice in the top-performing (20%) treatment centres, and provides insight into the features of practice that could lead to optimization of patient outcomes.

 Ooi, C. Y. & Durie, P. R. Cystic fibrosis transmembrane conductance regulator (CFTR) gene mutations in pancreatitis. *J. Cyst. Fibros.* 11, 355–362 (2012).

- 58. Ratjen, F. & Doring, G. Cystic fibrosis. *Lancet* **361**, 681–689 (2003).
- Rowe, S. M., Miller, S. & Sorscher, E. J. Cystic fibrosis N. Engl. J. Med. 352, 1992–2001 (2005).
- Boucher, R. C. Airway surface dehydration in cystic fibrosis: pathogenesis and therapy. *Annu. Rev. Med.* 58, 157–170 (2007).
- Adebamiro, A., Cheng, Y., Rao, U. S., Danahay, H. & Bridges, R. J. A segment of
 <u>'</u>ENaC mediates elastase activation of Na⁺ transport. *J. Gen. Physiol.* **130**, 611–629 (2007).
- Caldwell, R. A., Boucher, R. C. & Stutts, M. J. Neutrophil elastase activates near-silent epithelial Na⁺ channels and increases airway epithelial Na⁺ transport. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 288, L813–819 (2005).
- Passero, C. J. *et al.* TMPRSS4-dependent activation of the epithelial sodium channel requires cleavage of the γ-subunit distal to the furin cleavage site. *Am. J. Physiol. Renal Physiol.* **302**, F1–8 (2012).
- Matsui, H. *et al.* Evidence for periciliary liquid layer depletion, not abnormal ion composition, in the pathogenesis of cystic fibrosis airways disease. *Cell* **95**, 1005–1015 (1998).
- Button, B. *et al.* A periciliary brush promotes the lung health by separating the mucus layer from airway epithelia. *Science* 337, 937–941 (2012).
- Chen, J. H. *et al*. Loss of anion transport without increased sodium absorption characterizes newborn porcine cystic fibrosis airway epithelia. *Cell* 143, 911–923 (2010).
- Tuggle, K. L. et al. Characterization of defects in ion transport and tissue development in cystic fibrosis transmembrane conductance regulator (CFTR)knockout rats. *PLoS ONE* 9, e91253 (2014).
- Itani, O. A. *et al.* Human cystic fibrosis airway epithelia have reduced CI⁻ conductance but not increased Na⁺ conductance. *Proc. Natl Acad. Sci. USA* **108**, 10260–10265 (2011).
- Birket, S. E. *et al.* A functional anatomic defect of the cystic fibrosis airway. *Am. J. Respir. Crit. Care Med.* **190**, 421–432 (2014).
 Hoegger, M. J. *et al.* Cystic fibrosis. Impaired mucus
- Hoegger, M. J. et al. Cystic fibrosis. Impaired mucus detachment disrupts mucociliary transport in a piglet model of cystic fibrosis. Science 345, 818–822 (2014).
- Corcoran, T. E. *et al.* Absorptive clearance of DTPA as an aerosol-based biomarker in the cystic fibrosis airway. *Eur. Respir. J.* 35, 781–786 (2010).
- Schutte, A. *et al.* Microbial-induced meprin beta cleavage in MUC2 mucin and a functional CFTR channel are required to release anchored small intestinal mucus. *Proc. Natl Acad. Sci. USA* 111, 12396–12401 (2014).
- Henderson, A. G. *et al.* Cystic fibrosis airway secretions exhibit mucin hyperconcentration and increased osmotic pressure. *J. Clin. Invest.* **124**, 3047–3060 (2014).
- Quinton, P. M. Cystic fibrosis: impaired bicarbonate secretion and mucoviscidosis. *Lancet* **372**, 415–417 (2008).
- Garcia, M. A., Yang, N. & Quinton, P. M. Normal mouse intestinal mucus release requires cystic fibrosis transmembrane regulator-dependent bicarbonate serration. *J. Clin. Invest*. **119**, 2613–2622 (2009)
- secretion. J. Clin. Invest. 119, 2613–2622 (2009).
 Gustafsson, J. K. et al. Bicarbonate and functional CFTR channel are required for proper mucin secretion and link cystic fibrosis with its mucus phenotype. J. Exp. Med. 209, 1263–1272 (2012).
 References 70, 71 and 76 demonstrate that a defect in mucociliary clearance emanates from abnormalities of mucus in cystic fibrosis, including its adhesive properties, shown in mouse models in which the intestines are bicarbonate-dependent, porcine airway gland ducts submerged in saline and the human airway surface, even in the setting of adequate periciliary layer depth.
 Keiser, N. W. et al. Defective innate immunity and
- Keiser, N. W. et al. Defective innate immunity and hyper-inflammation in newborn CFTR-knockout ferret lungs. Am. J. Respir. Cell. Mol. Biol. 15 Oct 2014 [epub ahead of print].
- Rogers, C. S. *et al.* Disruption of the CFTR gene produces a model of cystic fibrosis in newborn pigs. *Science* 321, 1837–1841 (2008).
- Kunzelmann, K. & Mehta, A. CFTR: a hub for kinases and crosstalk of cAMP and Ca^{2+,} FEBS J. 280, 4417–4429 (2013).
- Garnett, J. P. et al. Novel role for pendrin in orchestrating bicarbonate secretion in cystic fibrosis transmembrane conductance regulator (CFTR)expressing airway serous cells. J. Biol. Chem. 286, 41069–41082 (2011).

- Pezzulo, A. A. *et al.* Reduced airway surface pH impairs bacterial killing in the porcine cystic fibrosis lung. *Nature* 487, 109–113 (2012).
- Pohl, K. et al. A neutrophil intrinsic impairment affecting Rab27a and degranulation in cystic fibrosis is corrected by CFTR potentiator therapy. *Blood* 124, 999–1009 (2014).
- Bruscia, E. M. *et al.* Macrophages directly contribute to the exaggerated inflammatory response in cystic fibrosis transmembrane conductance regulator⁻¹⁻ mice. *Am. J. Respir. Cell. Mol. Biol.* **40**, 295–304 (2009).
- Wang, X. *et al.* Hsp90 cochaperone Aha1 downregulation rescues misfolding of CFTR in cystic fibrosis. *Cell* **127**, 803–815 (2006).
- Bedrossian, C., Greenberg, S., Singer, D., Hansen, J. & Rosenberg, H. The lung in cystic fibrosis: a quantitative study including prevalence of pathologic findings among different age groups. *Hum. Pathol.* 7, 195–204 (1976).
- Livraghi-Butrico, A. *et al.* Mucus clearance, MyD88-dependent and MyD88-independent immunity modulate lung susceptibility to spontaneous bacterial infection and inflammation. *Mucosal Immunol.* 5, 397–408 (2012).
 Hybiske, K. *et al.* Effects of cystic fibrosis
- Hybiske, K. *et al.* Effects of cystic fibrosis transmembrane conductance regulator and Delta F508CFTR on inflammatory response, ER stress, and Ca²⁺ of airway epithelia. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **293**, 11250–11260 (2007)
- Mol. Physiol. 293, L1250–L1260 (2007).
 88. Mott, L. S. *et al.* Progression of early structural lung disease in young children with cystic fibrosis assessed using CT. *Thorax* 67, 509–516 (2011).
- Sly, P. D. *et al.* Risk factors for bronchiectasis in children with cystic fibrosis. *N. Engl. J. Med.* 368, 1963–1970 (2013).
 This article highlights that neutrophil elastase activity in bronchoalveolar lavage fluid in early life of children with cystic fibrosis undergoing bronchoscopy is associated with early bronchiectasis. This finding shows the importance of early detection of lung infection and inflammation, and the importance of its
- treatment.
 90. Gehrig, S. *et al.* Lack of neutrophil elastase reduces inflammation, mucus hypersecretion, and emphysema, but not mucus obstruction, in mice with cystic fibrosis-like lung disease. *Am. J. Respir. Crit. Care Med.* 189, 1082–1092 (2014).
- Vandivier, R. W. et al. Elastase-mediated phosphatidylserine receptor cleavage impairs apoptotic cell clearance in cystic fibrosis and bronchiectasis. J. Clin. Invest. 109, 661–670 (2002)
- bronchiectasis. J. Clin. Invest. 109, 661–670 (2002).
 Berger, M., Sorensen, R. U., Tosi, M. F., Dearborn, D. G. & Doring, G. Complement receptor expression on neutrophils at an inflammatory site, the *Pseudomonas*-infected lung in cystic fibrosis. J. Clin. Invest. 84, 1302–1313 (1989).
- J. Clin. Invest. 84, 1302–1313 (1989).
 93. Day, B. J., van Heeckeren, A. M., Min, E. & Velsor, L. W. Role for cystic fibrosis transmembrane conductance regulator protein in a glutathione response to bronchopulmonary pseudomonas infection. Infect. Immun 72, 2045–2051 (2004)
- Infection. Infect. Immun. **72**, 2045–2051 (2004).
 Roum, J. H., Buhl, R., McElvaney, N. G., Borok, Z. & Crystal, R. G. Systemic deficiency of glutathione in cystic fibrosis. *J. Appl. Physiol.* **75**, 2419–2424 (1993).
- Knowles, M. R. & Boucher, R. C. Mucus clearance as a primary innate defense mechanism for mammalian airways. J. Clin. Invest. 109, 571–577 (2002).
- Emerson, J., Rosenfeld, M., McNamara, S., Ramsey, B. & Gibson, R. L. *Pseudomonas aeruginosa* and other predictors of mortality and morbidity in young children with cystic fibrosis. *Pediatr. Pulmonol.* 34, 91–100 (2002).
- Nixon, G. M. *et al.* Clinical outcome after early *Pseudomonas aeruginosa* infection in cystic fibrosis. *J. Pediatr.* **138**, 699–704 (2001).
- Gangell, C. *et al.* Inflammatory responses to individual microorganisms in the lungs of children with cystic fibrosis. *Clin. Infect. Dis.* 53, 425–432 (2011).
- van Ewijk, B. E. *et al.* Prevalence and impact of respiratory viral infections in young children with cystic fibrosis: prospective cohort study. *Pediatrics* 122, 1171–1176 (2008).
- 100. Wat, D. *et al.* The role of respiratory viruses in cystic fibrosis. *J. Cyst. Fibros.* **7**, 320–328 (2008).
- Wark, P. A. *et al.* Viral infections trigger exacerbations of cystic fibrosis in adults and children. *Eur. Respir. J.* 40, 510–512 (2012).

- 102. Amin, R., Dupuis, A., Aaron, S. D. & Ratjen, F. The effect of chronic infection with *Aspergillus fumigatus* on lung function and hospitalization in patients with cystic fibrosis. *Chest* **137**, 171–176 (2010).
- Dasenbrook, E. C. *et al.* Association between respiratory tract methicillin-resistant *Staphylococcus aureus* and survival in cystic fibrosis. *JAMA* 303, 2386–2392 (2010).
- 104. Govan, J. R., Brown, A. R. & Jones, A. M. Evolving epidemiology of *Pseudomonas aeruginosa* and the *Burkholderia cepacia* complex in cystic fibrosis lung infection. *Future Microbiol.* 2, 153–164 (2007).
- 105. Ledson, M. J., Gallagher, M. J., Jackson, M., Hart, C. A. & Walshaw, M. J. Outcome of *Burkholderia cepacia* colonisation in an adult cystic fibrosis centre. *Thorax* 57, 142–145 (2002).
- Aaron, S. D. et al. Infection with transmissible strains of *Pseudomonas aeruginosa* and clinical outcomes in adults with cystic fibrosis. *JAMA* **304**, 2145–2153 (2010).
- 107. Armstrong, D. S. et al. Detection of a widespread clone of Pseudomonas aeruginosa in a pediatric cystic fibrosis clinic. Am. J. Respir. Crit. Care Med. 166, 983–987 (2002).
- McCallum, S. J. et al. Superinfection with a transmissible strain of *Pseudomonas aeruginosa* in adults with cystic fibrosis chronically colonised by *P. aeruginosa. Lancet* 358, 558–560 (2001).
- 109. Cramer, N. *et al.* Molecular epidemiology of chronic *Pseudomonas aeruginosa* airway infections in cystic fibrosis. *PLoS ONE* 7, e50731 (2012).
- 110. Kidd, T. J. et al. Pseudomonas aeruginosa exhibits frequent recombination, but only a limited association between genotype and ecological setting. *PLoS ONE* 7, e44199 (2012).
- Wiehlmann, L. et al. Effective prevention of Pseudomonas aeruginosa cross-infection at a cystic fibrosis centre — results of a 10-year prospective study. Int. J. Med. Microbiol. 302, 69–77 (2012).
- 112. Griffiths, A. L. *et al.* Effects of segregation on an epidemic *Pseudomonas aeruginosa* strain in a cystic fibrosis clinic. *Am. J. Respir. Crit. Care Med.* **171**, 1020–1025 (2005).
- 113. Adjemian, J., Olivier, K. N. & Prevots, D. R. Nontuberculous mycobacteria among patients with cystic fibrosis in the United States: screening practices and environmental risk. Am. J. Respir. Crit. Care Med. **190**, 581–586 (2014).
- 114. Qvist, T. et al. Epidemiology of nontuberculous mycobacteria among patients with cystic fibrosis in Scandinavia. J. Cyst. Fibros. 14, 46–52 (2014).
- 115. Bryant, J. M. et al. Whole-genome sequencing to identify transmission of Mycobacterium abscessus between patients with cystic fibrosis: a retrospective cohort study. Lancet 381, 1551–1560 (2013).
- 116. Rogers, G. B. et al. Bacterial diversity in cases of lung infection in cystic fibrosis patients: 16S ribosomal DNA (rDNA) length heterogeneity PCR and 16S rDNA terminal restriction fragment length polymorphism profiling. J. Clin. Microbiol. 41, 3548–3558 (2003).
- 117. Tunney, M. M. et al. Detection of anaerobic bacteria in high numbers in sputum from patients with cystic fibrosis. Am. J. Respir. Crit. Care Med. **177**, 995–1001 (2008).
- 118. LiPuma, J. J. Expanding our understanding of respiratory microbiota in cystic fibrosis. *Ann. Am. Thorac. Soc.* **11**, 1084–1085 (2014).
- Worlitzsch, D. et al. Effects of reduced mucus oxygen concentration in airway *Pseudomonas* infections of cystic fibrosis patients. J. Clin. Invest. **109**, 317–325 (2002).
- 120. Bragonzi, A. et al. Pseudomonas aeruginosa microevolution during cystic fibrosis lung infection establishes clones with adapted virulence. Am. J. Respir. Crit. Care Med. 180, 138–145 (2009).
- J. Respir. Crit. Care Med. 180, 138–145 (2009).
 121. Marvig, R. L., Sommer, L. M., Molin, S. & Johansen, H. K. Convergent evolution and adaptation of *Pseudomonas aeruginosa* within patients with cystic fibrosis. Nat. Genet. 47, 57–64 (2015).
- 122. Young, R. L. et al. Neutrophil extracellular trap (NET)-mediated killing of *Pseudomonas aeruginosa:* evidence of acquired resistance within the CF airway, independent of CFTR. *PLoS ONE* 6, e23637 (2011).
- 123. Regamey, N. *et al.* Distinct patterns of inflammation in the airway lumen and bronchial mucosa of children with cystic fibrosis. *Thorax* 67, 164–170 (2012).
- 124. Tan, H. L. *et al.* The Th17 pathway in cystic fibrosis lung disease. *Am. J. Respir. Crit. Care Med.* **184**, 252–258 (2011).

- 125. Hector, A. et al. Regulatory T cell impairment in cystic fibrosis patients with chronic *Pseudomonas* infection. *Am. J. Respir. Crit. Care Med.* **191**, 914–923 (2015).
- 126. Ooi, C. Y. et al. Does extensive genotyping and nasal potential difference testing clarify the diagnosis of cystic fibrosis among patients with single-organ manifestations of cystic fibrosis? *Thorax* 69, 254–260 (2014).

This manuscript looks at the use of ancillary testing in the diagnosis of cystic fibrosis. Despite increasing sophistication of ion transport measurements and genetic testing, the diagnosis frequently remains in doubt, underscoring the importance of clinical acumen in these indeterminate cases.

- 127. Ooi, C. Y. *et al.* Comparing the American and European diagnostic guidelines for cystic fibrosis: same disease, different language? *Thorax* 67, 618–624 (2012). The article discusses the nomenclature of and the approach to infants with an equivocal newborn screening test. This is a very difficult area of uncertainty for paediatricians and families, and this consensus statement, whether the nomenclature is thought to be correct or not, offers important guidance.
- 128. Stewart, B., Zabner, J., Shuber, A. P., Welsh, M. J. & McCray, P. B. Jr. Normal sweat chloride values do not exclude the diagnosis of cystic fibrosis. *Am. J. Respir. Crit. Care Med.* **151**, 899–903 (1995).
- 129. Knowles, M., Gatzy, J. & Boucher, R. Increased bioelectric potential difference across respiratory epithelia in cystic fibrosis. *N. Engl. J. Med.* **305**, 1489–1495 (1981).
- Ooi, C. Y. *et al.* Does integration of various ion channel measurements improve diagnostic performance in cystic fibrosis? *Ann. Am. Thorac Soc.* **11**, 562–570 (2014).
- Vermeulen, F., Proesmans, M., Boon, M. & De Boeck, K. Improved repeatability of nasal potential difference with a larger surface catheter. J. Cyst. Fibros. [online], http://dx.doi.org/10.1016/ i.icf.2014.08.006 (2014).
- j.jcf. 2014.08.006 (2014).
 132. Veeze, H. J., Sinaasappel, M., Bijman, J., Bouquet, J. & de Jonge, H. R. Ion transport abnormalities in rectal suction biopsies from children with cystic fibrosis. *Gastroenterology* 101, 398–403 (1991).
- 133. Cohen-Cymberknoh, M. *et al.* Evaluation of the intestinal current measurement method as a diagnostic test for cystic fibrosis. *Pediatr. Pulmonol.* 48, 229–235 (2013).
- Dekkers, J. F. et al. A functional CFTR assay using primary cystic fibrosis intestinal organoids. *Nat. Med.* 19, 939–945 (2013).
- 135. Quinton, P. et al. β-adrenergic sweat secretion as a diagnostic test for cystic fibrosis. Am. J. Respir. Crit. Care Med. 186, 732–739 (2012).
- 136. Lebecque, P. et al. Mutations of the cystic fibrosis gene and intermediate sweat chloride levels in children. Am. J. Respir. Crit. Care Med. 165, 757–761 (2002).
- 137. Bush, A. & Wallis, C. Time to think again: cystic fibrosis is not an 'all or none' disease. *Pediatr. Pulmonol.* **30**, 139–144 (2000).
- 138. Sims, E. J. *et al.* Economic implications of newborn screening for cystic fibrosis: a cost of illness retrospective cohort study. *Lancet* **369**, 1187–1195 (2007).
- 139. Farrell, P. M. *et al.* Nutritional benefits of neonatal screening for cystic fibrosis. Wisconsin Cyst. Fibrosis Neonatal Screen. *Study Group. N. Engl. J. Med.* 337, 963–969 (1997).
 This article is a major report on the only randomized controlled trial of screening. Screening led to improved nutritional outcomes but no respiratory improvements. This finding probably reflects suboptimal care in one clinic, making the important point that if cystic fibrosis newborn
- screening is instituted, the best standards of care for the screen-positive babies must be available. 140. Wilcken, B. & Gaskin, K. More evidence to favour newborn screening for cystic fibrosis. *Lancet* **369**,
- 1146–1147 (2007).
 141. Balfour-Lynn, I. M. Newborn screening for cystic fibrosis, evidence for boxofit. Arch. Dis. Child **93**, 7
- fibrosis: evidence for benefit. Arch. Dis. Child **93**, 7–10 (2008).
 142. Wagener, J. S., Zemanick, E. T. & Sontag, M. K.
- Newborn screening for cystic fibrosis. *Curr. Opin. Pediatr.* 24, 329–335 (2012).
 143. Ranganathan, S. C. *et al.* Airway function in infants
- Kanganathan, S. C. *et al.* Airway function in infants newly diagnosed with cystic fibrosis. *Lancet* 358, 1964–1965 (2001).

- 144. Ranganathan, S. C. *et al.* The evolution of airway function in early childhood following clinical diagnosis of cystic fibrosis. *Am. J. Respir. Crit. Care Med.* **169**, 928–933 (2004).
- 145. Southern, K. W. et al. A survey of newborn screening for cystic fibrosis in Europe. J. Cyst. Fibros. 6, 57–65 (2007).
- 146. Vernooij-van Langen, A. M. *et al.* Novel strategies in newborn screening for cystic fibrosis: a prospective controlled study. *Thorax* 67, 289–295 (2012).
- 147. Southern, K. W. Determining the optimal newborn screening protocol for cystic fibrosis. *Thorax* 67, 281–282 (2012).
- Roussey, M. et al. Neonatal screening of cystic fibrosis: diagnostic problems with CFTR mild mutations. J. Inherit Metab. Dis. 30, 613 (2007).
- 149. Scotet, V. et al. Immunoreactive trypsin/DNA newborn screening for cystic fibrosis: should the R117H variant be included in CFTR mutation panels? Pediatrics 118, e1523-e1529 (2006).
- O'Sullivan, B. P., Zwerdling, R. G., Dorkin, H. L., Comeau, A. M. & Parad, R. Early pulmonary manifestation of cystic fibrosis in children with the ΔF508/R117H-7T genotype. *Pediatrics* 118, 1260–1265 (2006).
- 151. Peckham, D., Conway, S. P., Morton, A., Jones, A. & Webb, K. Delayed diagnosis of cystic fibrosis associated with R117H on a background of 7T polythymidine tract at intron 8. *J. Cyst. Fibros.* 5, 63–65 (2006).
- 152. de Nooijer, R. A. *et al.* Assessment of CFTR function in homozygous R117H-7T subjects. *J. Cyst. Fibros.* **10**, 326–332 (2011).
- 153. Cystic Fibrosis, F. et al. Cystic Fibrosis Foundation practice guidelines for the management of infants with cystic fibrosis transmembrane conductance regulator-related metabolic syndrome during the first two years of life and beyond. J. Pediatr. 155, \$106-\$116 (2009).
- 154. Mayell, S. J. et al. A European consensus for the evaluation and management of infants with an equivocal diagnosis following newborn screening for cystic fibrosis, J. Cust. Fibros. 8, 71–78 (2009).
- Ren, C. L., Desai, H., Platt, M. & Dixon, M. Clinical outcomes in infants with cystic fibrosis transmembrane conductance regulator (CFTR) related metabolic syndrome. *Pediatr. Pulmonol.* 46, 1079–1084 (2011).
- 156. Wielputz, M. O. *et al.* Magnetic resonance imaging detects changes in structure and perfusion, and response to therapy in early cystic fibrosis lung disease. *Am. J. Respir. Crit. Care Med.* **189**, 956–965 (2014).
- 157. Aurora, P. *et al.* Multiple breath inert gas washout as a measure of ventilation distribution in children with cystic fibrosis. *Thorax* 59, 1068–1073 (2004).
- Aurora, P. et al. Multiple-breath washout as a marker of lung disease in preschool children with cystic fibrosis. Am. J. Respir. Crit. Care Med. 171, 249–256 (2005).
- 159. Kraemer, R., Blum, A., Schibler, A., Ammann, R. A. & Gallati, S. Ventilation inhomogeneities in relation to standard lung function in patients with cystic fibrosis. *Am J. Respir Crit Care Med* **171** 371–378 (2005)
- Am. J. Respir. Crit. Care Med. 171, 371–378 (2005).
 160. Lum, S. et al. Early detection of cystic fibrosis lung disease: multiple-breath washout versus raised volume tests. Thorax 62, 341–347 (2007).
- 161. Hoo, A. F. *et al.* Lung function is abnormal in 3-month-old infants with cystic fibrosis diagnosed by newborn screening. *Thorax* 67, 874–881 (2012).
- 162. Sly, P. D. et al. Lung disease at diagnosis in infants with cystic fibrosis detected by newborn screening. Am. J. Respir. Crit. Care Med. 180, 146–152 (2009).
- Pillarisetti, N. *et al.* Infection, inflammation, and lung function decline in infants with cystic fibrosis. *Am. J. Respir. Crit. Care Med.* **184**, 75–81 (2011).
 Nguyen, T. T. *et al.* Evolution of lung function during
- 164. Nguyen, T. T. et al. Evolution of lung function during the first year of life in newborn screened cystic fibrosis infants. Thorax 69, 910–917 (2013).
- 165. Brennan, L. C. *et al.* S7 Evolution of lung function during the first two years of life in infants with cystic fibrosis diagnosed by newborn screening. *Thorax* 68, A6–A7 (2013).
- 166. Amin, R. et al. The effect of dornase alfa on ventilation inhomogeneity in patients with cystic fibrosis. Eur. Respir. J. 37, 806–812 (2011).
- 167. Amin, R. et al. Hypertonic saline improves the LCI in paediatric CF patients with normal lung function. *Thorax* 65, 379–383 (2010).
- 168. Vermeulen, F., Proesmans, M., Boon, M., Havermans, T. & De Boeck, K. Lung clearance index

predicts pulmonary exacerbations in young patients with cystic fibrosis. *Thorax* **69**, 39–45 (2014).

- 169. Maisonneuve, P., Marshall, B. C., Knapp, E. A. & Lowenfels, A. B. Cancer risk in cystic fibrosis: a 20-year nationwide study from the United States. J. Natl Cancer Inst. 105, 122–129 (2013).
- Thia, L. P. et al. Is chest CT useful in newborn screened infants with cystic fibrosis at 1 year of age? *Thorax* 69, 320–327 (2014).
 Gustafsson, P. M., De Jong, P. A., Tiddens, H. A. &
- 171. Gustafsson, P. M., De Jong, P. A., Tiddens, H. A. & Lindblad, A. Multiple-breath inert gas washout and spirometry versus structural lung disease in cystic fibrosis. *Thorax* 63, 129–134 (2008).
- 172. Owens, C. M. *et al.* Lung Clearance Index and HRCT are complementary markers of lung abnormalities in young children with CF. *Thorax* 66, 481–488 (2011).
- Stafler, P., Davies, J. C., Balfour-Lynn, I. M., Rosenthal, M. & Bush, A. Bronchoscopy in cystic fibrosis infants diagnosed by newborn screening. *Pediatr. Pulmonol.* 46, 696–700 (2011).
 Wainwright, C. E. *et al.* Effect of bronchoalveolar
- 174. Wainwright, C. E. *et al.* Effect of bronchoalveolar lavage-directed therapy on *Pseudomonas aeruginosa* infection and structural lung injury in children with cystic fibrosis: a randomized trial. *JAMA* **306**, 163–171 (2011).
- 175. King, V. V. Upper respiratory disease, sinusitis, and polyposis. *Clin. Rev. Allergy* **9**, 143–157 (1991).
- Henriksson, G. *et al.* Nasal polyps in cystic fibrosis: clinical endoscopic study with nasal lavage fluid analysis. *Chest* **121**, 40–47 (2002).
 Hansen, S. K. *et al.* Evolution and diversification of
- 177. Hansen, S. K. et al. Evolution and diversification of *Pseudomonas aeruginosa* in the paranasal sinuses of cystic fibrosis children have implications for chronic lung infection. *ISME J.* 6, 31–45 (2012).
- Rubinstein, S., Moss, R. & Lewiston, N. Constipation and meconium ileus equivalent in patients with cystic fibrosis. *Pediatrics* **78**, 473–479 (1986).
 Blondeau, K. *et al.* Gastro-oesophageal reflux and
- 179. Blondeau, K. *et al.* Gastro-oesophageal reflux and aspiration of gastric contents in adult patients with cystic fibrosis. *Gut* **57**, 1049–1055 (2008).
- Ledder, O., Haller, W., Couper, R. T., Lewindon, P. & Oliver, M. Cystic fibrosis: an update for clinicians. Part 2: hepatobiliary and pancreatic manifestations. J. Gastroenterol. Hepatol. 29, 1954–1962 (2014).
- Borowitz, D. et al. Castrointestinal outcomes and confounders in cystic fibrosis. J. Pediatr. Gastroenterol. Nutr. 41, 273–285 (2005).
- Lewindon, P. J., Robb, T. A., Moore, D. J., Davidson, G. P. & Martin, A. J. Bowel dysfunction in cystic fibrosis: importance of breath testing. *J. Paediatr. Child Health* 34, 79–82 (1998).
- 183. Konstan, M. W. *et al.* Growth and nutritional indexes in early life predict pulmonary function in cystic fibrosis. J. Pediatr. **142**, 624–630 (2003).
- 184. Sharma, R. *et al.* Wasting as an independent predictor of mortality in patients with cystic fibrosis. *Thorax* 56, 746–750 (2001).
- Corbett, K. *et al.* Cystic fibrosis-associated liver disease: a population-based study. *J. Pediatr.* 145, 327–332 (2004).
- 186. Rowland, M. et al. Outcome in cystic fibrosis liver disease. Am. J. Gastroenterol. 106, 104–109 (2011).
- 187. Moran, A. *et al.* Clinical care guidelines for cystic fibrosis-related diabetes: a position statement of the American Diabetes Association and a clinical practice guideline of the Cystic Fibrosis Foundation, endorsed by the Pediatric Endocrine Society. *Diabetes Care* 33, 2697–2708 (2010).
- 188. Lewis, C. *et al.* Diabetes-related mortality in adults with cystic fibrosis. Role of genotype and sex. *Am. J. Respir. Crit. Care Med.* **191**, 194–200 (2015).
- Neinstein, L. S., Stewart, D., Wang, C. I. & Johnson, I. Menstrual dysfunction in cystic fibrosis. *J. Adolesc. Health Care* 4, 153–157 (1983).
- 190. Stern, M., Wiedemann, B., Wenzlaff, P. & German Cystic Fibrosis Quality Assessment, G. From registry to quality management: the German Cystic Fibrosis Quality Assessment project 1995–2006. *Eur. Respir.* J. **31**, 29–35 (2008).
- 191. Marshall, B. C. & Nelson, E. C. Accelerating implementation of biomedical research advances: critical elements of a successful 10 year Cystic Fibrosis Foundation healthcare delivery improvement initiative. *BMJ Qual. Saf.* 23 (Suppl. 1), i95–i103 (2014).
- 192. McCormick, J. *et al.* Comparative demographics of the European cystic fibrosis population: a cross-sectional database analysis. *Lancet* **375**, 1007–1013 (2010).
- 193. Colombo, C. & Littlewood, J. The implementation of standards of care in Europe: state of the art. J. Cyst. Fibros. 10 (Suppl. 2), S7–S15 (2011).

- 194. Stern, M., Niemann, N., Wiedemann, B., Wenzlaff, P. & German, C. G. Benchmarking improves quality in cystic fibrosis care: a pilot project involving 12 centres. *Int. J. Qual. Health Care* 23, 349–356 (2011).
- 195. Schechter, M. S. Benchmarking to improve the quality of cystic fibrosis care. *Curr. Opin. Pulm. Med.* 18, 596–601 (2012).
- 196. Conway, S. *et al.* European Cystic Fibrosis Society standards of care: framework for the Cystic Fibrosis Centre. *J. Cyst. Fibros.* **13**, S3–S22 (2014).
- European Cystic Fibrosis Society. ECFS Patient Registry annual data report 2010. (European Cystic Fibrosis Society, 2010).
- 198. Cystic Fibrosis Trust. UK CF Registry annual report 2011. (Cystic Fibrosis Trust, 2013).
- 199. Goss, C. H. *et al.* Children and young adults with CF in the USA have better lung function compared with the UK. *Thorax* **70**, 229–236 (2014).
- Cystic Fibrosis Australia. 15th Annual Report from the Cystic Fibrosis Data Registry. (Cystic Fibrosis Australia, 2013).
- Stern, M. *et al.* European Cystic Fibrosis Society standards of care: quality management in cystic fibrosis. *J. Cyst. Fibros.* **13**, S43–S59 (2014).
- Quittner, A. L. *et al.* Pulmonary medication adherence and health-care use in cystic fibrosis. *Chest* 146, 142–151 (2014).
- Abbott, J., Dodd, M. & Webb, A. K. Health perceptions and treatment adherence in adults with cystic fibrosis. *Thorax* 51, 1233–1238 (1996).
- 204. Conway, S. P., Pond, M. N., Hamnett, T. & Watson, A. Compliance with treatment in adult patients with cystic fibrosis. *Thorax* 51, 29–33 (1996).
- Plant, B. J., Goss, C. H., Plant, W. D. & Bell, S. C. Management of comorbidities in older patients with cystic fibrosis. *Lancet Respir. Med.* 1, 164–174 (2013).
- Grasemann, H. & Ratjen, F. Early lung disease in cystic fibrosis. *Lancet Respir. Med.* 1, 148–157 (2013).
- Flume, P. A. *et al.* Cystic fibrosis pulmonary guidelines: treatment of pulmonary exacerbations. *Am. J. Respir. Crit. Care Med.* 180, 802–808 (2009).
- 208. Smyth, A. R. *et al.* European Cystic Fibrosis Society Standards of Care: Best Practice guidelines. *J. Cyst. Fibros.* **13**, S23–S42 (2014).
- Okumura, M. J. *et al.* Improving transition from paediatric to adult cystic fibrosis care: programme implementation and evaluation. *BMJ Qual. Saf.* 23 (Suppl. 1), i64–i72 (2014).
- Lester, M. K. & Flume, P. A. Airway-clearance therapy guidelines and implementation. *Respir. Care* 54, 733–753 (2009).
 McIlwaine, M. P. *et al.* Long-term multicentre
- Mcllwaine, M. P. et al. Long-term multicentre randomised controlled study of high frequency chest wall oscillation versus positive expiratory pressure mask in cystic fibrosis. *Thorax* 68, 746–751 (2013).
- 212. Schneiderman, J. E. *et al.* Longitudinal relationship between physical activity and lung health in patients with cystic fibrosis. *Eur. Respir. J.* **43**, 817–823 (2014).
- 213. Cox, N. S., Alison, J. A. & Holland, A. E. Interventions for promoting physical activity in people with cystic fibrosis. *Cochrane Database Syst. Rev.* **12**, CD009448 (2013).
- Fuchs, H. J. et al. Effect of aerosolized recombinant human DNase on exacerbations of respiratory symptoms and on pulmonary function in patients with cystic fibrosis. N. Engl. J. Med. 331, 637–642 (1994)
- cystic fibrosis. N. Engl. J. Med. 331, 637–642 (1994).
 215. Jones, A. P. & Wallis, C. E. Recombinant human deoxyribonuclease for cystic fibrosis. Cochrane Database Syst. Rev. 3, CD001127 (2003).
- 216. Shah, P. I. *et al.* Recombinant human DNase I in cystic fibrosis patients with severe pulmonary disease: a short-term, double-blind study followed by six months open-label treatment. *Eur. Respir. J.* 8, 954–958 (1995).
- Quan, J. M. et al. A two-year randomized, placebocontrolled trial of dornase alfa in young patients with cystic fibrosis with mild lung function abnormalities. J. Pediatr. 139, 813–820 (2001).
- 218. Flume, P. A. *et al.* Cystic fibrosis pulmonary guidelines: chronic medications for maintenance of lung health. *Am. J. Respir. Crit. Care Med.* **176**, 957–969 (2007). References **197** and **218** describe current standards of care of implementing a cystic fibrosis centre, and for the management of patients with chronic maintenance therapies, respectively.
- Elkins, M. R. *et al.* A controlled trial of long-term inhaled hypertonic saline in patients with cystic fibrosis. *N. Engl. J. Med.* 354, 229–240 (2006).

- Rosenfeld, M. *et al.* Inhaled hypertonic saline in infants and children younger than 6 years with cystic fibrosis: the ISIS randomized controlled trial. *JAMA* **307**, 2269–2277 (2012).
- 221. Subbarao, P. et al. Lung clearance index as an outcome measure for clinical trials in young children with cystic fibrosis. A pilot study using inhaled hypertonic saline. Am. J. Respir. Crit. Care Med. 188, 456–460 (2013).
- Bilton, D. *et al.* Inhaled dry powder mannitol in cystic fibrosis: an efficacy and safety study. *Eur. Respir. J.* 38, 1071–1080 (2011).
- Aitken, M. L. *et al.* Long-term inhaled dry powder mannitol in cystic fibrosis: an international randomized study. *Am. J. Respir. Crit. Care Med.* 185, 645–652 (2012).
- 224. Ramsey, B. W. *et al.* Efficacy of aerosolized tobramycin in patients with cystic fibrosis. *N. Engl. J. Med.* **328**, 1740–1746 (1993).
- 225. Ramsey, B. W. *et al.* Intermittent administration of inhaled tobramycin in patients with cystic fibrosis. *N. Engl. J. Med.* **340**, 23–30 (1999).
- Konstan, M. W. *et al.* Safety, efficacy and convenience of tobramycin inhalation powder in cystic fibrosis patients: The EAGER trial. *J. Cyst. Fibros.* **10**, 54–61 (2011).
- McCoy, K. S. et al. Inhaled aztreonam lysine for chronic airway Pseudomonas aeruginosa in cystic fibrosis. Am. J. Respir. Crit. Care Med. **178**, 921–928 (2008).
- Retsch-Bogart, G. Z. et al. A phase 2 study of aztreonam lysine for inhalation to treat patients with cystic fibrosis and *Pseudomonas aeruginosa* infection. *Pediatr. Pulmonol.* 43, 47–58 (2008).
- 229. Doring, G., Flume, P., Heijerman, H., Elborn, J. S. & Consensus Study, G. Treatment of lung infection in patients with cystic fibrosis: current and future strategies. J. Cyst. Fibros. 11, 461–479 (2012).
- Tullis, D. E. *et al.* Inhaled aztreonam for chronic Burkholderia infection in cystic fibrosis: a placebocontrolled trial. J. Cyst. Fibros. 13, 296–305 (2014).
- 231. Wolter, J. *et al*. Effect of long term treatment with azithromycin on disease parameters in cystic fibrosis: a randomised trial. *Thorax* **57**, 212–216 (2002).
- 232. Saiman, L. *et al.* Azithromycin in patients with cystic fibrosis chronically infected with *Pseudomonas aeruginosa*: a randomized controlled trial. *JAMA* 290, 1749–1756 (2003).
- 233. Saiman, L. *et al.* Effect of azithromycin on pulmonary function in patients with cystic fibrosis uninfected with *Pseudomonas aeruginosa*: a randomized controlled trial. JAMA 305, 1707–1715 (2010).
- 234. Clement, A. *et al.* Long term effects of azithromycin in patients with cystic fibrosis: A double blind, placebo controlled trial. *Thorax* **61**, 895–902 (2006).
- Konstan, M. W., Byard, P. J., Hoppel, C. L. & Davis, P. B. Effect of high-dose ibuprofen in patients with cystic fibrosis. *N. Engl. J. Med.* **332**, 848–854 (1995).
- Lands, L. C. & Stanojevic, S. Oral non-steroidal antiinflammatory drug therapy for lung disease in cystic fibrosis. *Cochrane Database Syst, Rev.* 6, CD001505 (2013).
- 237. Leeboh, H. S., Williams, M., Kirkpatrick, J. A. & Colten, H. R. Alternate-day prednisone reduces morbidity and improves pulmonary function in cystic fibrosis. *Lancet* 2, 686–688 (1985).
- Eigen, H., Rosenstein, B. J., FitzSimmons, S. & Schidlow, D. V. A multicenter study of alternate-day prednisone therapy in patients with cystic fibrosis. *J. Pediatr.* **126**, 515–523 (1995).
- Balfour-Lynn, I. M. & Welch, K. Inhaled corticosteroids for cystic fibrosis. *Cochrane Database Syst. Rev.* 11, CD001915 (2009).
- Balfour-Lynn, I. M. et al. Multicenter randomized controlled trial of withdrawal of inhaled corticosteroids in cystic fibrosis. Am. J. Respir. Crit. Care Med. 173, 1356–1362 (2006).
- 241. Konstan, M. W. et al. A randomized double blind, placebo controlled phase 2 trial of BIIL 284 BS (an LTB4 receptor antagonist) for the treatment of lung disease in children and adults with cystic fibrosis. J. Cyst. Fibros. 13, 148–155 (2014).
- Hirche, T. O. *et al.* Practical guidelines: lung transplantation in patients with cystic fibrosis. *Pulm. Med.* 2014, 621342 (2014).
- Med. 2014, 621342 (2014).
 243. Madden, B. P. et al. Noninvasive ventilation in cystic fibrosis patients with acute or chronic respiratory failure. *Eur. Respir. J.* 19, 310–313 (2002).
- 244. Gill, D. R. & Hyde, S. C. Delivery of genes into the CF airway. *Thorax* **69**, 962–964 (2014).

- 245. Crystal, R. G. *et al*. Administration of an adenovirus containing the human CFTR cDNA to the respiratory tract of individuals with cystic fibrosis. *Nat. Genet.* 8, 42–51 (1994).
- 246. Harvey, B. G., Hackett, N. R., Ely, S. & Crystal, R. G. Host responses and persistence of vector genome following intrabronchial administration of an E1-E3adenovirus gene transfer vector to normal individuals. *Mol. Ther.* **3**, 206–215 (2001).
- 247. Knowles, M. R. et al. A controlled study of adenoviral-vector-mediated gene transfer in the nasal epithelium of patients with cystic fibrosis. N. Engl. J. Med. 333, 823–831 (1995).
- Moss, R. B. *et al.* Repeated adeno-associated virus serotype 2 aerosol-mediated cystic fibrosis transmembrane regulator gene transfer to the lungs of patients with cystic fibrosis: a multicenter, doubleblind, placebo-controlled trial. *Chest* **125**, 509–521 (2004).
 Moss, R. B. *et al.* Repeated aerosolized AAV-CFTR for
- 249. Moss, R. B. *et al.* Repeated aerosolized AAV-CFTR for treatment of cystic fibrosis: a randomized placebocontrolled phase 2B trial. *Hum. Gene Ther.* 18, 726–732 (2007).
- 250. Griesenbach, U. et al. Assessment of F/ HN-pseudotyped lentivirus as a clinically relevant vector for lung gene therapy. Am. J. Respir. Crit. Care Med. 186, 846–856 (2012).
- 251. Alton, E. W. et al. A randomised, double-blind, placebo-controlled phase IIB clinical trial of repeated application of gene therapy in patients with cystic fibrosis. *Thorax* 68, 1075–1077 (2013).
- 252. Bangel-Ruland, N. *et al.* Cystic fibrosis transmembrane conductance regulator-mRNA delivery: a novel alternative for cystic fibrosis gene therapy. J. Gene Med. **15**, 414–426 (2013).
- 253. Wilschanski, M. *et al.* Gentamicin-induced correction of CFTR function in patients with cystic fibrosis and CFTR stop mutations. *N. Engl. J. Med.* **349**, 1433–1441 (2003).
- 254. Linde, L. *et al.* Nonsense-mediated mRNA decay affects nonsense transcript levels and governs response of cystic fibrosis patients to gentamicin. *J. Clin. Invest.* **117**, 683–692 (2007).
- 255. Kerem, E. *et al.* Ataluren for the treatment of nonsense-mutation cystic fibrosis: a randomised, double-blind, placebo-controlled phase 3 trial. *Lancet Respir. Med.* 2, 539–547 (2014).
- Lancet Respir. Med. 2, 539–547 (2014). 256. Study of ataluren (PTC124®) in cystic fibrosis [online], https://clinicaltrials.gov/ct2/show/ NCT02107859 (2014).
- 257. Van Goor, F. et al. Rescue of CF airway epithelial cell function in vitro by a CFTR potentiator, VX-770. Proc. Natl Acad. Sci. USA 106, 18825–18830 (2009).
- 258. Rowe, S. M. et al. Clinical mechanism of the cystic fibrosis transmembrane conductance regulator potentiator ivacaftor in G551D-mediated cystic fibrosis. Am. J. Respir. Crit. Care Med. **190**, 1775–184 (2014).
- 259. De Boeck, K. et al. Efficacy and safety of ivacaftor in patients with cystic fibrosis and a non-G551D gating mutation. J. Cyst. Fibros. 13, 674–680 (2014).
- 260. Sloane, P. A. *et al.* A pharmacologic approach to acquired cystic fibrosis transmembrane conductance regulator dysfunction in smoking related lung disease. *PLoS ONE* 7, e39809 (2012).
- Flume, P. A. *et al.* Ivacaftor in subjects with cystic fibrosis who are homozygous for the F508del-CFTR mutation. *Chest* **142**, 718–724 (2012).
 Boyle, M. P. *et al.* A CFTR corrector (lumacaftor) and
- 262. Boyle, M. P. et al. A CFTR corrector (lumacaftor) and a CFTR potentiator (ivacaftor) for treatment of patients with cystic fibrosis who have a phe508del CFTR mutation: a phase 2 randomised controlled trial 2, 527–538 (2014).

- 263. Van Goor, F. et al. Correction of the F508del-CFTR protein processing defect *in vitro* by the investigational drug VX-809. Proc. Natl Acad. Sci. USA 108, 18843–18848 (2011).
- 264. Veit, G. *et al.* Some gating potentiators, including VX-770, diminish ΔF508–CFTR functional expression. *Sci. Transl. Med.* 6, 246ra97 (2014).
- Sci. Transl. Med. 6, 246ra97 (2014).
 265. Cholon, D. M. et al. Potentiator ivacaftor abrogates pharmacological correction of ΔF508 CFTR in cystic fibrosis. Sci. Transl. Med. 6, 246ra96 (2014).
- Okiyoneda, T. *et al.* Mechanism-based corrector combination restores DeltaF508-CFTR folding and function. *Nat. Chem. Biol.* 9, 444–454 (2013).
- Accurso, F. J. *et al.* Denufosol tetrasodium in patients with cystic fibrosis and normal to mildly impaired lung function. *Am. J. Respir. Crit. Care Med.* 183, 627–634 (2011).
- Ratjen, F. *et al.* Long term effects of denufosol tetrasodium in patients with cystic fibrosis. *J. Cyst. Fibros.* 11, 539–549 (2012).
- Burrows, E. F., Southern, K. W. & Noone, P. G. Sodium channel blockers for cystic fibrosis. *Cochrane Database Syst, Rev.* 4, CD005087 (2014).
- 270. Quittner, A. L., Alpern, A. N. & Kimberg, C. I. in Kendig and Chernick's Disorders of the Respiratory Tract in Children. 8th edn (eds Wilmott, R. W., Boat, T. F., Bush, A., Chernick, V., Deterding, R. R. & Ratjen, F.) 251–260 (Elsevier (Saunders), 2012).
- 271. Goss, C. H., Edwards, T. C., Ramsey, B. W., Aitken, M. L. & Patrick, D. L. Patient-reported respiratory symptoms in cystic fibrosis. *J. Cyst. Fibros.* 8, 245–252 (2009).
- Schmidt, A. M. et al. Exercise and quality of life in patients with cystic fibrosis: A 12-week intervention study. *Physiother Theory Pract.* 27, 548–556 (2011).
- 273. Schechter, M. S. *et al.* Long-term effects of pregnancy and motherhood on disease outcomes of women with cystic fibrosis. *Ann. Am. Thorac Soc.* **10**, 213–219 (2013).
- 274. Sawicki, G. S. *et al.* Longitudinal assessment of healthrelated quality of life in an observational cohort of patients with cystic fibrosis. *Pediatr. Pulmonol.* 46, 36–44 (2011).
- 275. Snyder, C. F. *et al.* Implementing patient-reported outcomes assessment in clinical practice: a review of the options and considerations. *Qual. Life Res.* **21**, 1305–1314 (2012).
- 276. Sawicki, G. S., Sellers, D. E. & Robinson, W. M. High treatment burden in adults with cystic fibrosis: challenges to disease self-management. J. Cyst. Fibros. 8, 91–96 (2009).
- Quittner, A. L. Measurement of quality of life in cystic fibrosis. *Curr. Opin. Pulm. Med.* 4, 326–331 (1998).
 Goss, C. H. & Quittner, A. L. Patient-reported
- Goss, C. H. & Quittner, A. L. Patient-reported outcomes in cystic fibrosis. *Proc. Am. Thorac. Soc.* 4, 378–386 (2007).
- Quittner, A. L. *et al.* Erratum to: Psychometric evaluation of the Cystic Fibrosis Questionnaire-Revised in a national, US sample. *Qual. Life Res.* 21, 1279–1290 (2012).
- Donaldson, S. H. *et al.* Mucus clearance and lung function in cystic fibrosis with hypertonic saline. *N. Engl. J. Med.* **354**, 241–250 (2006).
- Sawicki, G. S. *et al.* Treatment complexity in cystic fibrosis: trends over time and associations with sitespecific outcomes. *J. Cyst. Fibros.* **12**, 461–467 (2013).

References 277 and 281 show the treatment burden and the complexity in patients with cystic fibrosis, respectively.

 Quittner, A. L. *et al.* Impact of socioeconomic status, race, and ethnicity on quality of life in patients with cystic fibrosis in the United States. *Chest* **137**, 642–650 (2010).

- Quittner, A. L. *et al.* Prevalence of depression and anxiety in patients with cystic fibrosis and parent caregivers: results of The International Depression Epidemiological Study across nine countries. *Thorax* 69, 1090–1097 (2014).
- 284. Smith, B. A., Modì, A. C., Quittner, A. L. & Wood, B. L. Depressive symptoms in children with cystic fibrosis and parents and its effects on adherence to airway clearance. *Pediatr. Pulmonol.* **45**, 756–763 (2010).
- 285. Riekert, K. A., Bartlett, S. J., Boyle, M. P., Krishnan, J. A. & Rand, C. S. The association between depression, lung function, and health-related quality of life among adults with cystic fibrosis. *Chest* **132**, 231–237 (2007).
- Goldbeck, L. *et al.* Prevalence of symptoms of anxiety and depression in German patients with cystic fibrosis. *Chest* **138**, 929–936 (2010).
- Snell, C., Fernandes, S., Bujoreanu, I. S. & Garcia, G. Depression, illness severity, and healthcare utilization in cystic fibrosis. *Pediatr. Pulmonol.* 49, 1177–1181 (2014).
- Wong, A. P. *et al.* Directed differentiation of human pluripotent stem cells into mature airway epithelia expressing functional CFTR protein. *Nat. Biotech.* **30**, 876–882 (2012).
 References 135 and 288 describe model systems to assess response to CFTR modulators *in vitro* by either using intestinal organoids or skin-derived pluripotent stem cells transformed into airway epithelial cells, respectively.

Author contributions

Introduction (F.R.); Epidemiology (C.H.G.), Mechanisms/ pathophysiology (S.M.R. and S.C.B.); Diagnosis, screening and prevention (A.B., F.R. and S.M.R.); Management (S.C.B. and F.R.); Quality of life (A.L.Q.); Outlook (F.R. and A.B.); overview of the Primer (F.R.).

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